Screening Assessment for the Challenge

C.I. Pigment Yellow 34

Chemical Abstracts Service Registry Number 1344-37-2

Environment Canada Health Canada

November 2008

Synopsis

Pursuant to section 74 of the *Canadian Environmental Protection Act, 1999* (CEPA 1999), the Ministers of the Environment and of Health have conducted a screening assessment of C.I. Pigment Yellow 34, Chemical Abstracts Service Registry Number (CAS RN) 1344-37-2. The substance C.I. Pigment Yellow 34 was identified in the categorization of the Domestic Substances List as a high priority for action under the Ministerial Challenge. The substance was identified as a high priority because it was considered to pose greatest potential for exposure (GPE) to individuals in Canada and had been classified by other agencies on the basis of carcinogenicity, reproductive toxicity and developmental toxicity. The substance also met the ecological categorization criteria for persistence and inherent toxicity to aquatic organisms. Therefore, this assessment of C.I. Pigment Yellow 34 focuses on information relevant to the evaluation of both human health and ecological risks.

In response to a notice issued under section 71 of CEPA 1999, in 2006 C.I. Pigment Yellow 34 was reported to be manufactured in and imported into Canada. After exports, the amount remaining for use in this country ranged between 1 000 000 and 10 000 000 kg. It is primarily used for plastic formulation for commercial applications and export; commercial, non-consumer paints and coatings; and commercial printing inks or coatings used for plastics and certain outdoor applications such as commercial identification decals.

There were no empirical data identified regarding measured concentrations of C.I. Pigment Yellow 34 in environmental media (i.e., air, water, soil and food) in Canada. Given the physical and chemical properties and sources of this substance, exposure to C.I. Pigment Yellow 34 is expected to be negligible via drinking water, ambient air or consumer products. Exposure to the general population in Canada is expected to be predominantly from soils, although these exposures are expected to be low due to the primarily commercial use of the substance, very limited industrial releases, and the encapsulation and incorporation of the substance into a solid matrix. However, these exposures could not be quantified due to lack of measured concentrations.

The substance C.I. Pigment Yellow 34 is considered persistent because it contains metal ions, lead (Pb²⁺) and the chromate (CrO₄²⁻) ions, which are considered to be infinitely persistent. Therefore, C.I. Pigment Yellow 34 meets the persistence criteria as set out in the *Persistence and Bioaccumulation Regulations*. The current state of the science does not allow for the unambiguous interpretation of the bioaccumulation potential of metal-containing inorganic substances such as C.I. Pigment Yellow 34. Experimental toxicity studies suggest that the substance is not hazardous to aquatic organisms at a loading rate (100 mg/L) that is considered to represent a reasonable environmental worst-case scenario. Additionally, considering its low solubility, it is unlikely that organisms associated with other compartments would be harmed by exposure to this substance.

Based principally on the weight of evidence based classification of C.I. Pigment Yellow 34 by the European Commission, and the assessment of hexavalent chromium and inorganic lead compounds by several national and international agencies, a critical effect for the

characterization of risk to human health is carcinogenicity. The substance C.I. Pigment Yellow 34, together with lead chromate and C.I. Pigment Red 104, was carcinogenic in rats after subcutaneous and intramuscular administration and these animal studies are supported by epidemiological studies, which indicate an increased frequency of lung cancer in chromate pigment production workers. As well, C.I. Pigment Yellow 34 or its principal components were genotoxic in a limited number of *in vitro* and *in vivo* experimental systems.

On the basis of the carcinogenicity of C.I. Pigment Yellow 34, for which there may be a probability of harm at any level of exposure, it is concluded that C.I. Pigment Yellow 34 is a substance that may be entering the environment in a quantity or concentration or under conditions that constitute or may constitute a danger in Canada to human life or health.

On the basis of ecological hazard and reported releases of C.I. Pigment Yellow 34, it is concluded that this substance is not entering the environment in a quantity or concentration or under conditions that have or may have an immediate or long-term harmful effect on the environment or its biological diversity, or that constitute or may constitute a danger to the environment on which life depends.

In addition and where relevant, research and monitoring will support verification of assumptions used during the screening assessment and, where appropriate, the performance of potential control measures identified during the risk management phase.

Based on the information available, it is concluded that C.I. Pigment Yellow 34 meets one or more of the criteria set out in section 64 of the *Canadian Environmental Protection Act*, 1999.

Introduction

The Canadian Environmental Protection Act, 1999 (CEPA 1999) (Canada 1999) requires the Minister of the Environment and the Minister of Health to conduct screening assessments of substances that have met the categorization criteria set out in the Act to determine whether these substances present or may present a risk to the environment or human health. Based on the results of a screening assessment, the Ministers can propose to take no further action with respect to the substance, to add the substance to the Priority Substances List (PSL) for further assessment, or to recommend that the substance be added to the List of Toxic Substances in Schedule 1 of the Act and, where applicable, the implementation of virtual elimination.

Based on the information obtained through the categorization process, the Ministers identified a number of substances as high priorities for action. These include substances that

- met all of the ecological categorization criteria, including persistence (P), bioaccumulation potential (B) and inherent toxicity to aquatic organisms (iT), and were believed to be in commerce; and/or
- met the categorization criteria for greatest potential for exposure (GPE) or presented an intermediate potential for exposure (IPE), and had been identified as posing a high hazard to human health based on classifications by other national or international agencies for carcinogenicity, genotoxicity, developmental toxicity or reproductive toxicity.

The Ministers therefore published a notice of intent in the *Canada Gazette*, Part I, on December 9, 2006 (Canada 2006), that challenged industry and other interested stakeholders to submit, within specified timelines, specific information that may be used to inform risk assessment and to develop and benchmark best practices for the risk management and product stewardship of those substances identified as high priorities.

The substance C.I. Pigment Yellow 34 was identified as a high priority for assessment of human health risk because it was considered to present GPE and had been classified by other agencies on the basis of carcinogenicity, reproductive toxicity and developmental toxicity. The Challenge for C.I. Pigment Yellow 34 was published in the *Canada Gazette* on May 12, 2007 (Canada 2007). A substance profile was released at the same time. The substance profile presented the technical information available prior to December 2005 that formed the basis for categorization of this substance. As a result of the Challenge, submissions of information were received.

Although C.I. Pigment Yellow 34 was determined to be a high priority for assessment with respect to risks to human health, it also met the ecological categorization criteria for persistence and inherent toxicity for aquatic organisms during the categorization of the Domestic Substances List. Therefore, this assessment focuses on information relevant to the evaluation of human health and ecological risks.

Under CEPA 1999, screening assessments focus on information critical to determining whether a substance meets the criteria for defining a chemical as toxic as set out in section 64 of the Act, where

"64. [...] a substance is toxic if it is entering or may enter the environment in a quantity or concentration or under conditions that

- (a) have or may have an immediate or long-term harmful effect on the environment or its biological diversity;
- (b) constitute or may constitute a danger to the environment on which life depends; or
- (c) constitute or may constitute a danger in Canada to human life or health."

Screening assessments examine scientific information and develop conclusions by incorporating a weight of evidence approach and precaution.

This screening assessment includes consideration of information on chemical properties, hazards, uses and exposure, including the additional information submitted under the Challenge. Data relevant to the screening assessment of this substance were identified in original literature, review and assessment documents, stakeholder research reports and from recent literature searches, up to October 2007 for the environmental section, and up to January 2008 for the health effects section of the document. Key studies were critically evaluated; modelling results may have been used to reach conclusions.

Evaluation of risk to human health involves consideration of data relevant to estimation of exposure (non-occupational) of the general population, as well as information on health hazards (based principally on the weight of evidence assessments of other agencies that were used for prioritization of the substance). Decisions for human health are based on the nature of the critical effect and/or margins between conservative effect levels and estimates of exposure, taking into account confidence in the completeness of the identified databases on both exposure and effects, within a screening context. The screening assessment does not represent an exhaustive or critical review of all available data. Rather, it presents a summary of the critical information upon which the proposed conclusion is based.

This screening assessment was prepared by staff in the Existing Substances Programs at Health Canada and Environment Canada and incorporates input from other programs within these departments. This assessment has undergone external written peer review/consultation. Comments on the technical portions relevant to human health were received from scientific experts selected and directed by Toxicology Excellence for Risk Assessment (TERA), including Donna Vorhees (The Science Collaborative), Glenn Talaska (University of Cincinnati) and Joan Strawson (TERA). Comments on these sections were also received from Herman Gibb, Sciences International. While external comments were taken into consideration, the final content and outcome of the screening risk assessment remain the responsibility of Health Canada and Environment Canada. Additionally, the draft of this screening assessment was subject to a 60-day public comment period. The critical information and considerations upon which the assessment is based are summarized below.

Substance Identity

The use of the common name C.I. Pigment Yellow 34 in this assessment strictly refers to the substance identified as CAS RN 1344-37-2, even though this name could designate a mixture (Colour Index International, 2002).

Table 1. Substance identity

Registry Number (CAS RN) Name on Domestic Substances List (DSL) Pigment Yellow 34 (TSCA, AICS, PICCS, ASIA-PAC, NZIoC); Lead sulfochromate yellow; (EINECS); Pigment Yellow 34 (ECL); Mix-Crystal Lead Sulfochromate-Molybdate (PICCS); Chrome Yellow (PICCS); Chrome Yellow (PICCS); Chrome Yellow (PICCS); Chrome Yellow Lead Sulfochromate-Molybdate (PICCS); Chrome Yellow Lead Sulfochromate (PICCS) C 103; C 103; (pigment); 77600; 77603; C.P. Chrome Yellow Light 1066; Chrome Yellow Light 1074; Chrome Yellow Medium 1074; Chrome Yellow Medium 1085; Chrome Yellow Medium 1298; Chromastral Green Y; Chrome orange; Chrome Yellow 10G; Chrome Yellow 4G; Chrome Yellow 4GIL tight; Chrome Yellow 500LSG; Chrome Yellow 6G; Chrome Yellow GG; Chrome Yellow 500LSG; Chrome Yellow GG; Chrome Yellow Wight; Chrome Yellow 62E; Chrome Yellow FA; Chrome Yellow Might; Chrome Yellow Lemon; Chrome Yellow LF AA; Chrome Yellow Medium; Chrome Yellow Lemon; Chrome Yellow Indelie; Chrome Yellow Medium Y 469D); Chrome yellow middle; Chrome Yellow Modium Y 469D); Chrome yellow; Horna Chrome Yellow Primrose; Chromium yellow; Dainichi Chrome Yellow 10G; Dainichi Chrome Yellow Pigment GMN 35; Chrome Yellow Primrose; Chromium yellow; Dainichi Chrome Yellow 10G; Dainichi Chrome Yellow Yellow Yellow Wight; Chrome Yellow Yellow; Dainichi Chrome Yellow; Pigment GMN 35; Chrome Yellow Primrose; Chromium yellow; Dainichi Chrome Yellow; Porthome Yellow Aark GL 35; Horna Chrome Yellow GUH 41; Horna Chrome Yellow Primrose; Chrome Yellow; Pigment Yellow; Pigment GMN 325; Light chrome yellow; Medium chrome yellow; Middle chrome; Middle Chrome BHG; Middle chrome Yellow; Perma Yellow 150S); Perma Yellow 5G; Primrose chrome; Primrose Yellow; Pure Lemon Chrome L 3G; Pure Primrose Chrome L 3G; Pure Primrose Chrome L 10G; Pure Primrose	Chemical Abstracts Service				
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Chrome Yellow 4G; Chrome Yellow 4GL Light; Chrome Yellow 500LSG; Chrome Yellow 5G; Chrome Yellow 5GF; Chrome Yellow 62E; Chrome Yellow 6GL Primrose; Chrome Yellow A 241; Chrome Yellow G; Chrome Yellow GL Medium; Chrome Yellow Lemon; Chrome Yellow LF AA; Chrome yellow light; Chrome Yellow Light Y 434D; Chrome yellow medium; Chrome Yellow Medium Y 469D; Chrome yellow middle; Chrome Yellow NEO 5GS; Chrome Yellow Pigment GMN 35; Chrome Yellow Primrose; Chromium yellow; Dainichi Chrome Yellow 10G; Dainichi Chrome Yellow 5G; Dark chrome yellow; Horna Chrome Yellow dark GL 35; Horna Chrome Yellow GUH 41; Horna Chrome Yellow medium GU 25; Krolor Yellow KY 787D; Krolor Yellow KY 788D; KZh 2; KZh 3; KZh 3 (pigment); Lead sulphochromate; Lemon Chrome A 3G; Lemon Chrome C 4G; Lemon Chrome Yellow 325; Light chrome yellow; Medium chrome yellow; Middle chrome; Middle Chrome BHG; Middle chrome yellow; Perma Yellow 1650S; Perma Yellow 5G; Primrose chrome; Primrose yellow; Pure Lemon Chrome 13GS; Pure Lemon Chrome 13GS; Pure Lemon Chrome L 3GS; Pure Middle Chrome L 3GS; Pure Middle Chrome 24883; Pure Middle Chrome LG; Pure Primrose Chrome 24880; Pure Primrose Chrome 24881; Pure Primrose Chrome L					
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62E; Chrome Yellow 6GL Primrose; Chrome Yellow A 241; Chrome Yellow G; Chrome Yellow GL Medium; Chrome Yellow Lemon; Chrome Yellow LF AA; Chrome yellow light; Chrome Yellow Light Y 434D; Chrome yellow medium; Chrome Yellow Medium Y 469D; Chrome yellow middle; Chrome Yellow NEO 5GS; Chrome Yellow Pigment GMN 35; Chrome Yellow Primrose; Chromium yellow; Dainichi Chrome Yellow 10G; Dainichi Chrome Yellow 5G; Dark chrome yellow; Horna Chrome Yellow dark GL 35; Horna Chrome Yellow GUH 41; Horna Chrome Yellow medium GU 25; Krolor Yellow KY 787D; Krolor Yellow KY 788D; KZh 2; KZh 3; KZh 3 (pigment); Lead sulphochromate; Lemon Chrome A 3G; Lemon Chrome C 4G; Lemon Chrome Yellow; Middle chrome; Middle Chrome BHG; Middle chrome yellow; Middle chrome; Middle Chrome BHG; Middle chrome yellow; Perma Yellow 1650S; Perma Yellow 5G; Primrose chrome; Primrose yellow; Pure Lemon Chrome 24882; Pure Lemon Chrome L 3G; Pure Lemon Chrome L 3G; Pure Middle Chrome 24883; Pure Middle Chrome LG; Pure Primrose Chrome L 2480; Pure Primrose Chrome L		~			
Yellow G; Chrome Yellow GL Medium; Chrome Yellow Lemon; Chrome Yellow LF AA; Chrome yellow light; Chrome Yellow Light Y 434D; Chrome yellow medium; Chrome Yellow Medium Y 469D; Chrome yellow middle; Chrome Yellow NEO 5GS; Chrome Yellow Pigment GMN 35; Chrome Yellow Primrose; Chromium yellow; Dainichi Chrome Yellow 10G; Dainichi Chrome Yellow 5G; Dark chrome yellow; Horna Chrome Yellow dark GL 35; Horna Chrome Yellow GUH 41; Horna Chrome Yellow medium GU 25; Krolor Yellow KY 787D; Krolor Yellow KY 788D; KZh 2; KZh 3; KZh 3 (pigment); Lead sulphochromate; Lemon Chrome A 3G; Lemon Chrome C 4G; Lemon Chrome Yellow 325; Light chrome yellow; Medium chrome yellow; Middle chrome; Middle Chrome BHG; Middle chrome yellow; Perma Yellow 1650S; Perma Yellow 5G; Primrose chrome; Primrose yellow; Pure Lemon Chrome L 3G; Pure Lemon Chrome L 3G; Pure Lemon Chrome HL 3G; Pure Lemon Chrome L 3G; Pure Lemon Chrome L 3GS; Pure Middle Chrome 24883; Pure Middle Chrome LG; Pure Primrose Chrome 24880; Pure Primrose Chrome 24881; Pure Primrose Chrome L					
Chrome Yellow LF AA; Chrome yellow light; Chrome Yellow Light Y 434D; Chrome yellow medium; Chrome Yellow Medium Y 469D; Chrome yellow middle; Chrome Yellow NEO 5GS; Chrome Yellow Pigment GMN 35; Chrome Yellow Primrose; Chromium yellow; Dainichi Chrome Yellow 10G; Dainichi Chrome Yellow 5G; Dark chrome yellow; Horna Chrome Yellow dark GL 35; Horna Chrome Yellow GUH 41; Horna Chrome Yellow medium GU 25; Krolor Yellow KY 787D; Krolor Yellow KY 788D; KZh 2; KZh 3; KZh 3 (pigment); Lead sulphochromate; Lemon Chrome A 3G; Lemon Chrome C 4G; Lemon Chrome Yellow 325; Light chrome yellow; Medium chrome yellow; Middle chrome; Middle Chrome BHG; Middle chrome yellow; Perma Yellow 1650S; Perma Yellow 5G; Primrose chrome; Primrose yellow; Pure Lemon Chrome 24882; Pure Lemon Chrome L 3G; Pure Lemon Chrome L 3G; Pure Middle Chrome 24883; Pure Middle Chrome LG; Pure Primrose Chrome L 24880; Pure Primrose Chrome L4881; Pure Primrose Chrome L					
Y 434D; Chrome yellow medium; Chrome Yellow Medium Y 469D; Chrome yellow middle; Chrome Yellow NEO 5GS; Chrome Yellow Pigment GMN 35; Chrome Yellow Primrose; Chromium yellow; Dainichi Chrome Yellow 10G; Dainichi Chrome Yellow 5G; Dark chrome yellow; Horna Chrome Yellow dark GL 35; Horna Chrome Yellow GUH 41; Horna Chrome Yellow medium GU 25; Krolor Yellow KY 787D; Krolor Yellow KY 788D; KZh 2; KZh 3; KZh 3 (pigment); Lead sulphochromate; Lemon Chrome A 3G; Lemon Chrome C 4G; Lemon Chrome Yellow 325; Light chrome yellow; Medium chrome yellow; Middle chrome; Middle Chrome BHG; Middle chrome yellow; Perma Yellow 1650S; Perma Yellow 5G; Primrose chrome; Primrose yellow; Pure Lemon Chrome 24882; Pure Lemon Chrome 3GN; Pure Lemon Chrome HL 3G; Pure Lemon Chrome L 3G; Pure Lemon Chrome L 3GS; Pure Middle Chrome 24883; Pure Middle Chrome LG; Pure Primrose Chrome 24880; Pure Primrose Chrome 24881; Pure Primrose Chrome L					
Chrome yellow middle; Chrome Yellow NEO 5GS; Chrome Yellow Pigment GMN 35; Chrome Yellow Primrose; Chromium yellow; Dainichi Chrome Yellow 10G; Dainichi Chrome Yellow 5G; Dark chrome yellow; Horna Chrome Yellow dark GL 35; Horna Chrome Yellow GUH 41; Horna Chrome Yellow medium GU 25; Krolor Yellow KY 787D; Krolor Yellow KY 788D; KZh 2; KZh 3; KZh 3 (pigment); Lead sulphochromate; Lemon Chrome A 3G; Lemon Chrome C 4G; Lemon Chrome Yellow 325; Light chrome yellow; Medium chrome yellow; Middle chrome; Middle Chrome BHG; Middle chrome yellow; Perma Yellow 1650S; Perma Yellow 5G; Primrose chrome; Primrose yellow; Pure Lemon Chrome 24882; Pure Lemon Chrome A 3G; Pure Lemon Chrome L 3G; Pure Lemon Chrome L 3G; Pure Middle Chrome 24883; Pure Middle Chrome LG; Pure Primrose Chrome L 24880; Pure Primrose Chrome 24881; Pure Primrose Chrome L					
Other names Pigment GMN 35; Chrome Yellow Primrose; Chromium yellow; Dainichi Chrome Yellow 10G; Dainichi Chrome Yellow 5G; Dark chrome yellow; Horna Chrome Yellow dark GL 35; Horna Chrome Yellow GUH 41; Horna Chrome Yellow medium GU 25; Krolor Yellow KY 787D; Krolor Yellow KY 788D; KZh 2; KZh 3; KZh 3 (pigment); Lead sulphochromate; Lemon Chrome A 3G; Lemon Chrome C 4G; Lemon Chrome Yellow 325; Light chrome yellow; Medium chrome yellow; Middle chrome; Middle Chrome BHG; Middle chrome yellow; Perma Yellow 1650S; Perma Yellow 5G; Primrose chrome; Primrose yellow; Pure Lemon Chrome 24882; Pure Lemon Chrome 3GN; Pure Lemon Chrome HL 3G; Pure Lemon Chrome L 3G; Pure Lemon Chrome L 3GS; Pure Middle Chrome 24883; Pure Middle Chrome LG; Pure Primrose Chrome 24880; Pure Primrose Chrome L					
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Other names chrome yellow; Horna Chrome Yellow dark GL 35; Horna Chrome Yellow GUH 41; Horna Chrome Yellow medium GU 25; Krolor Yellow KY 787D; Krolor Yellow KY 788D; KZh 2; KZh 3; KZh 3 (pigment); Lead sulphochromate; Lemon Chrome A 3G; Lemon Chrome C 4G; Lemon Chrome Yellow 325; Light chrome yellow; Medium chrome yellow; Middle chrome; Middle Chrome BHG; Middle chrome yellow; Perma Yellow 1650S; Perma Yellow 5G; Primrose chrome; Primrose yellow; Pure Lemon Chrome 24882; Pure Lemon Chrome A 3G; Pure Lemon Chrome L 3G; Pure Middle Chrome 24883; Pure Middle Chrome LG; Pure Primrose Chrome L 24880; Pure Primrose Chrome L		·			
Yellow GUH 41; Horna Chrome Yellow medium GU 25; Krolor Yellow KY 787D; Krolor Yellow KY 788D; KZh 2; KZh 3; KZh 3 (pigment); Lead sulphochromate; Lemon Chrome A 3G; Lemon Chrome C 4G; Lemon Chrome Yellow 325; Light chrome yellow; Medium chrome yellow; Middle chrome; Middle Chrome BHG; Middle chrome yellow; Perma Yellow 1650S; Perma Yellow 5G; Primrose chrome; Primrose yellow; Pure Lemon Chrome 24882; Pure Lemon Chrome A 3G; Pure Lemon Chrome L 3G; Pure Middle Chrome 24883; Pure Middle Chrome LG; Pure Primrose Chrome L 24880; Pure Primrose Chrome L		Dainichi Chrome Yellow 10G; Dainichi Chrome Yellow 5G; Dark			
Yellow GUH 41; Horna Chrome Yellow medium GU 25; Krolor Yellow KY 787D; Krolor Yellow KY 788D; KZh 2; KZh 3; KZh 3 (pigment); Lead sulphochromate; Lemon Chrome A 3G; Lemon Chrome C 4G; Lemon Chrome Yellow 325; Light chrome yellow; Medium chrome yellow; Middle chrome; Middle Chrome BHG; Middle chrome yellow; Perma Yellow 1650S; Perma Yellow 5G; Primrose chrome; Primrose yellow; Pure Lemon Chrome 24882; Pure Lemon Chrome 3GN; Pure Lemon Chrome HL 3G; Pure Lemon Chrome L 3G; Pure Middle Chrome 24883; Pure Middle Chrome LG; Pure Primrose Chrome L 24880; Pure Primrose Chrome Chrome L	Other names	·			
(pigment); Lead sulphochromate; Lemon Chrome A 3G; Lemon Chrome C 4G; Lemon Chrome Yellow 325; Light chrome yellow; Medium chrome yellow; Middle chrome; Middle Chrome BHG; Middle chrome yellow; Perma Yellow 1650S; Perma Yellow 5G; Primrose chrome; Primrose yellow; Pure Lemon Chrome 24882; Pure Lemon Chrome 3GN; Pure Lemon Chrome HL 3G; Pure Lemon Chrome L 3G; Pure Lemon Chrome L 3GS; Pure Middle Chrome 24883; Pure Middle Chrome LG; Pure Primrose Chrome L 24880; Pure Primrose Chrome L	Curer marries	Yellow GUH 41; Horna Chrome Yellow medium GU 25; Krolor			
Chrome C 4G; Lemon Chrome Yellow 325; Light chrome yellow; Medium chrome yellow; Middle chrome; Middle Chrome BHG; Middle chrome yellow; Perma Yellow 1650S; Perma Yellow 5G; Primrose chrome; Primrose yellow; Pure Lemon Chrome 24882; Pure Lemon Chrome 3GN; Pure Lemon Chrome HL 3G; Pure Lemon Chrome L 3G; Pure Lemon Chrome L 3GS; Pure Middle Chrome 24883; Pure Middle Chrome LG; Pure Primrose Chrome L 24880; Pure Primrose Chrome L		Yellow KY 787D; Krolor Yellow KY 788D; KZh 2; KZh 3; KZh 3			
Medium chrome yellow; Middle chrome; Middle Chrome BHG; Middle chrome yellow; Perma Yellow 1650S; Perma Yellow 5G; Primrose chrome; Primrose yellow; Pure Lemon Chrome 24882; Pure Lemon Chrome 3GN; Pure Lemon Chrome HL 3G; Pure Lemon Chrome L 3G; Pure Lemon Chrome L 3GS; Pure Middle Chrome 24883; Pure Middle Chrome LG; Pure Primrose Chrome 24880; Pure Primrose Chrome 24881; Pure Primrose Chrome L		(pigment); Lead sulphochromate; Lemon Chrome A 3G; Lemon			
Middle chrome yellow; Perma Yellow 1650S; Perma Yellow 5G; Primrose chrome; Primrose yellow; Pure Lemon Chrome 24882; Pure Lemon Chrome 3GN; Pure Lemon Chrome HL 3G; Pure Lemon Chrome L 3G; Pure Lemon Chrome L 3GS; Pure Middle Chrome 24883; Pure Middle Chrome LG; Pure Primrose Chrome 24880; Pure Primrose Chrome L		Chrome C 4G; Lemon Chrome Yellow 325; Light chrome yellow;			
Primrose chrome; Primrose yellow; Pure Lemon Chrome 24882; Pure Lemon Chrome 3GN; Pure Lemon Chrome HL 3G; Pure Lemon Chrome L 3G; Pure Lemon Chrome L 3GS; Pure Middle Chrome 24883; Pure Middle Chrome LG; Pure Primrose Chrome 24880; Pure Primrose Chrome 24881; Pure Primrose Chrome L		Medium chrome yellow; Middle chrome; Middle Chrome BHG;			
Pure Lemon Chrome 3GN; Pure Lemon Chrome HL 3G; Pure Lemon Chrome L 3G; Pure Lemon Chrome L 3GS; Pure Middle Chrome 24883; Pure Middle Chrome LG; Pure Primrose Chrome 24880; Pure Primrose Chrome 24881; Pure Primrose Chrome L		Middle chrome yellow; Perma Yellow 1650S; Perma Yellow 5G;			
Lemon Chrome L 3G; Pure Lemon Chrome L 3GS; Pure Middle Chrome 24883; Pure Middle Chrome LG; Pure Primrose Chrome 24880; Pure Primrose Chrome 24881; Pure Primrose Chrome L		Primrose chrome; Primrose yellow; Pure Lemon Chrome 24882;			
Chrome 24883; Pure Middle Chrome LG; Pure Primrose Chrome 24880; Pure Primrose Chrome 24881; Pure Primrose Chrome L		Pure Lemon Chrome 3GN; Pure Lemon Chrome HL 3G; Pure			
24880; Pure Primrose Chrome 24881; Pure Primrose Chrome L		Lemon Chrome L 3G; Pure Lemon Chrome L 3GS; Pure Middle			
		Chrome 24883; Pure Middle Chrome LG; Pure Primrose Chrome			
10G; Pure Primrose Chrome L 6G; Renol Chrome Yellow Y 2G;		24880; Pure Primrose Chrome 24881; Pure Primrose Chrome L			
		10G; Pure Primrose Chrome L 6G; Renol Chrome Yellow Y 2G;			
Renol Chrome Yellow Y 2RS; Resino Yellow NSR 107; Sicomin		Renol Chrome Yellow Y 2RS; Resino Yellow NSR 107; Sicomin			
Yellow L 1122; Sicomin Yellow L 1630S; Sicomin Yellow L 1635;		Yellow L 1122; Sicomin Yellow L 1630S; Sicomin Yellow L 1635;			

	Supra Lemon Chrome 4Gl; Supra Lemon Chrome H 4G; Supra			
	Middle Chrome G; Supra Primrose Chrome 6G			
Chemical group (DSL stream)	UVCBs (<u>U</u> nknown or <u>V</u> ariable Composition, <u>C</u> omplex Reaction			
Chemical group (DSL stream)	Products, or <u>B</u> iological Materials) - Inorganics			
Chemical sub-group	Group IVA and group VIB element compounds; chromium VI-			
Chemical sub-group	containing; lead II-containing; oxides; sulfates			
Chemical formula	PbCrO ₄ * PbSO ₄ (Solid solution crystal of PbCrO ₄ and PbSO ₄ in			
Chemical formula	varying proportions) (Environment Canada 2007a)			
Chemical structure N/A				
Simplified Molecular Input				
Line Entry System (SMILES)	N/A			
Molecular mass	PbCrO ₄ = 323.2 g/mol (main constituent); PbSO ₄ = 303.3 g/mol			

Source: National Chemical Inventories (NCI). 2007: AICS (Australian Inventory of Chemical Substances); ASIA-PAC (Combined Inventories from the Asia-Pacific Region); ECL (Korean Existing Chemicals List); EINECS (European Inventory of Existing Chemical Substances); ELINCS (European List of Notified Chemical Substances), ENCS (Japanese Existing and New Chemical Substances); NZIoC (New Zealand Inventory of Chemicals); PICCS (Philippine Inventory of Chemicals and Chemical Substances); and TSCA (Toxic Substances Control Act Chemical Substance Inventory).

Physical and Chemical Properties

Table 2 contains experimental and modelled physical and chemical properties of C.I. Pigment Yellow 34 that are relevant to its environmental fate.

Table 2. Physical and chemical properties for C.I. Pigment Yellow 34

Property	Туре	Value (SI units) ¹	Temperature (C°)	Reference
Physical state	Experimental	Solid, yellow powder	(C')	Study Submission 2006c
Average particle size	Experimental	~ 200	-	Environment Canada 2007a
(μm)	Analogue ²	0.1-1	-	Lewis 1988
Melting point (°C)	Experimental	844	-	Lide 2006
Boiling point (°C)	-	Not available	-	-
		3800–6000 (3.8–6 g/cm ³)	20	IUCLID 2000
Density (kg/m ³)	Experimental	(6.12 g/cm^3)		Lide 2006
		5200–5400 (5.2–5.4 g/cm ³)		Study Submission 2006c
Vapour pressure (Pa)	Experimental	Negligible	Not indicated	IUCLID 2000
Henry's Law constant (Pa m³/mol)	Calculated	Negligible	-	-
Log K _{ow} (Octanol-water partition coefficient, dimensionless)	-	Not applicable	-	-
,	Experimental, parent substance	< 0.01	20	IUCLID 2000
	Experimental,	0.058	25	Weast 1965
	PbCrO ₄ (major component)	0.17	20	Lide 2006
Water solubility (mg/L)	Experimental, PbSO ₄ (minor component)	42.5	25	NICNAS 2007
	Experimental, total dissolved chromium ³	0.012; 0.10; 0.179	21–25; 19.5– 20.4; 13–17	Study Submission 2006a; 2006b; 2006c
	Experimental, total dissolved lead ²	0.02; 0.36; 0.223	21–25; 19.5– 20.4; 13–17	Study Submission 2006a; 2006b; 2006c
	Calculated, parent substance ⁴	0.062; 0.693; 0.764	21–25; 19.5– 20.4; 13–17	Study Submission 2006a; 2006b; 2006c
Log K _{oc} (Organic carbon-water partition coefficient, dimensionless)	-	Not applicable	-	-

Property	Туре	Value (SI units) ¹	Temperature (C°)	Reference
Log K _{sw} (Partition coefficient soil-water, dimensionless) ⁵	Experimental	Pb (3.444–5.212) Cr (2.812–4.764)	-	Thibault et al. 1990; Janssen et al. 1997; Sauvé et al. 2000
Log K _{sdw} (Partition coefficient sedimentwater, dimensionless) ⁵	Experimental	Pb (4.553–6.075) Cr (4.928–5.627)	-	Smock 1983; Timmermans et al. 1989; Keenan and Alikhan 1991; Van Hattum et al. 1991; Davis et al. 1996; Besser et al. 2001; Allison and Allison 2005
Log K _{ssw} (Partition coefficient suspended sediment-water, dimensionless) ⁵	Experimental	Pb (5.391–6.879) Cr (4.376–5.759)	-	Chiffoleau et al. 1994; Lofts and Tipping 2000; Roditi et al. 2000; Allison and Allison 2005; Gobeil et al. 2005

¹ Values in brackets represent the values in the original units as reported by the authors.

C.I. Pigment Yellow 34 contains principally lead chromate (PbCrO₄) and lead sulfate (PbSO₄). The proportion of these individual moieties in the pigment must be considered when evaluating exposure to each of the constituent metals. The *Pigment Handbook* (Lewis 1988) provides a range of composition percentages for the different constituents that are included in the second column of Table 3. Using these data with the molecular weights, the weight fractions of lead and chromate moieties are calculated for each constituent. Table 4 provides the total weight composition for each moiety by summing the contributions from each constituent.

Table 3. Composition range and weight fractions for C.I. Pigment Yellow 34

Constituent	Composition range	Average composition	Molecular weight (g/mol)		Weight fraction (%)	
	(%)	(%)	Pb	Other	Pb	Other
PbCrO ₄	61–76	68	207.2	116	44	24
PbSO ₄	20–38	29	207.2	96	20	9
Other	1-8					

² This is original data taken from C.I. Pigment Red 104 (CAS RN 12656-85-8); the particle size range for C.I. Pigment Yellow 34 is assumed to be similar.

 $^{^3}$ Based upon dissolution of the parent substance C.I. Pigment Yellow 34 after 18 to 24 hours of stirring in dilution test water (pH 7.1 to 8.4, room temperature), 0.2 or 0.45 μ m filtration and measurement of total dissolved metal in filtrate. The loading rate was 100 to 1000 mg of parent substance per litre.

⁴ Solubility of the parent substance was back-calculated using the total dissolved concentrations of the metals (Cr, Pb) and information on the composition of that parent substance.

⁵ Partition coefficients apply to the dissolved metal fractions.

Table 4. Weight fraction of specific moieties for C.I. Pigment Yellow 34

Moiety	Composition (%)
Pb	64
CrO ₄	24
SO ₄	9

Sources

The substance C.I. Pigment Yellow 34 is not known to be naturally produced in the environment. The principal metallic components of this substance, lead and chromium, are naturally occurring and as such are considered infinitely persistent. Lead concentrations in the rock of the upper continental crust have been determined to range between 17 and 20 ppm; chromium concentrations have been determined to be approximately 35 ppm (Reimann and de Caritat 1998).

As indicated in Table 2, these compounds are not highly soluble. However, while lead sulfate is present in the pigment at two-fold lower quantities, it is orders of magnitude more soluble. Therefore, lead sulfate may be a more significant source of dissolved lead from C.I. Pigment Yellow 34, despite being present in smaller quantities. However, there are various grades of pigments including those in which the pigment is encapsulated in a dense amorphous silica coating, which significantly reduces its solubility and bioavailability (Lewis 1988).

Based on a survey conducted under section 71 of CEPA 1999, in 2006 C.I. Pigment Yellow 34 was both manufactured in and imported into Canada (Environment Canada 2007b). Based on an exportation rate of around 75% of all substance manufactured (Environment Canada 2007a), between 1 000 000 and 10 000 000 kg of this substance would be remaining for use in this country.

Uses

According to the Color Pigments Manufacturers Association, the significant applications for these pigments in Canada are plastic formulation for commercial applications and export; commercial, non-consumer paints and coatings; and a very limited number of commercial printing inks or coatings used for plastics and certain outdoor applications such as commercial identification decals. For example, these pigments are used for applications that require safety attributes such as high visibility and so are used in traffic paint striping for highways and airports, and safety identification paints on buses, ambulances and fire trucks. Industrial paints using lead chromate pigments include automotive finishes, industrial and agricultural equipment, industrial baking enamels and air-dried finishes (Environment Canada 2007a).

The substance C.I. Pigment Yellow 34 is not used in consumer paints because the Canadian *Hazardous Products Act* prohibits furniture and other articles for children that are painted with a surface coating material that contains lead compounds of which the total lead content is more than 600 mg/kg (Canada 2005a). A concentration greater than 600 mg/kg would be required technically to manufacture a paint coloured with this substance (Environment Canada 2007a). The *Hazardous Products Act* also prohibits toys, equipment and other products for use by a child in learning or play and pencils and artists' brushes that have had a surface coating material applied to them that contains more than 600 mg/kg of total lead (Canada 2005a).

The Surface Coating Materials Regulations of the Hazardous Products Act (Canada 2005b) prohibits surface coatings (paints) containing greater than 600 mg/kg of total lead except

- 1. as an anti-corrosive or an anti-weathering coating applied to buildings or equipment used for agricultural or industrial purposes;
- 2. as an anti-corrosive or an anti-weathering coating applied to any structure, other than a building, used for agricultural, industrial or public purposes;
- 3. as a touch-up coating for metal surfaces;
- 4. on traffic signs;
- 5. for graphic art on billboards or similar displays;
- 6. for identification marks in industrial buildings; and
- 7. material for the purposes of arts, crafts or hobbies, other than material for use by children.

These regulations do not apply in the case of a surface coating material that is used exclusively in an industrial setting, other than for a use described [above], on surfaces where the finish is so durable and tightly adhered to the surface that it cannot be chewed or chipped under foreseeable use.

The substance C.I. Pigment Yellow 34 is not listed in the Food Additive Tables of Division 16 of the Food and Drug Regulations, and is therefore not permitted as a food colour in Canada. In addition, personal communication with the Health Protection and Foods Branch of Health Canada confirmed that this substance is not currently being used in food packaging material (as per email from the Food Packaging and Incidental Additives Section, Health Products and Food Branch, Health Canada, dated 2008 March 07, unreferenced).

Releases to the Environment

Submissions from a survey conducted under section 71 of CEPA 1999, in 2006 indicated that industrial releases of C.I. Pigment Yellow 34 to ambient air, water and soil were extremely low (i.e., less than 0.1% of total quantities manufactured in or imported into Canada) (Environment Canada 2007b). As well, there are provincial and municipal regulations or bylaws regarding levels of acceptable releases of lead or chromium (such as the *Ontario Regulation 419/05 Air Pollution – Local Air Quality* (Ontario 2005) and the Sewer Use By-Law (Durham Region 2004)).

Mass flow tool

To estimate potential release of the substance to the environment at all stages of its life cycle. a mass flow tool was used (Table 5). Empirical data concerning releases of this substance to the environment are not available. Therefore, for each identified type of use of the substance, the proportion and quantity of release to the different environmental media are estimated, as is the proportion of the substance chemically transformed or sent for waste disposal. Assumptions and input parameters used in making these estimates are based on information obtained from a variety of sources including responses to regulatory surveys, Statistics Canada, manufacturers' websites and technical databases. Of particular relevance are emission factors, which are generally expressed as the fraction of a substance released to the environment, particularly during its manufacture, formulation and use associated with industrial processes. Sources of such information include emission scenario documents, often developed under the auspices of the Organisation for Economic Cooperation and Development (OECD), and default assumptions used by different international chemical regulatory agencies. It is noted that the level of uncertainty in the mass of substance in circulation and quantity released to the environment generally increase towards the end of the life cycle.

Table 5. Estimated releases and losses of C.I. Pigment Yellow 34 to environmental media, transformation and distribution to management processes, based on the mass flow tool¹

le stage involved ²
strial use
e, waste disposal
e, formulation,
e and service life
e disposal
disposal

¹ For C.I. Pigment Yellow 34, information from the following OECD emission scenario documents was used to estimate releases to the environment and distribution of the substance, as summarized in this table: OECD 2004; Brooke and Crookes 2007. Values presented for releases to environmental media do not account for possible mitigation measures that may be in place in some locations (e.g., partial removal by sewage treatment plants). Specific assumptions used in derivation of these estimates are summarized in Environment Canada 2007c. ² Applicable stage(s): production-formulation-industrial use-consumer use-service life of article/product-waste disposal.

The results suggest that C.I. Pigment Yellow 34 mainly ends up in waste management sites (62.7%), due to the eventual disposal of manufactured items containing it. Of the substance, 1.9% is transformed, which in this case means destruction or modification of the structure of the substance during its incineration. Based largely on information contained in OECD emission scenario documents for manufacture, formulation and uses associated with this type of substance, it is estimated that 35.3% of C.I. Pigment Yellow 34 may be released to water. Negligible releases are expected to soil, groundwater and air. Although results from a survey conducted under section 71 of CEPA 1999, in 2006 (Environment Canada 2007b) indicate that releases to all media from industrial manufacture and formulation were extremely low (i.e., less than 0.1% of total manufactured or imported into Canada), the mass flow tool

results indicate that specific applications and/or post-application releases (e.g., from commercial use) are expected to make the greatest contribution to environmental levels.

Based on the above, water is expected to be the medium receiving the greatest proportion of Pigment Yellow 34 emitted during product manufacturing, formulation, industrial use and service life. It is anticipated that the majority of the substance bound in the product will be sent to landfills or incinerators for disposal. Note that the tool assumes no releases from such waste disposal sites.

Although no information is available on the quantity of importation of finished products containing C.I. Pigment Yellow 34, it is anticipated that the relative quantities released to the various environmental media would not be greatly different from those estimated here. However, it is likely that the percentage sent for waste management would be somewhat higher if importation of finished products were taken into consideration, and the percentage released to water would be lower.

Given some potentially dispersive uses of this chemical in specific coloured products, it is expected that C.I. Pigment Yellow 34 is being released into the Canadian environment mainly as a result of industrial use and service life of these products.

Environmental Fate

Solubility and dissociation

Inorganic metal-containing compounds often dissolve, dissociate and release ions into solution (Environment Canada 2003). The measured and estimated solubility in water of C.I. Pigment Yellow 34 is quite low ranging from < 0.01 to 0.764 mg/L (Table 2) with geometric and arithmetic means of 0.135 and 0.382 mg/L respectively. A relatively low proportion of the parent substance is thus expected to dissolve, dissociate and release the lead (Pb²⁺) and chromate (CrO₄²⁻) ions in typical aquatic media with a pH between 6 and 8 under conditions that are moderately oxic (~ 0.4–0.7 V, or dissolved oxygen > 4 mg/L). These moieties, released from the parent substance, are of greatest toxicological significance and are thus the moieties of concern for this assessment (Environment Canada 2003). In addition to the low solubility of the parent substance itself, its encapsulation of the substance in paints, plastics and coatings that are made to last for long periods of time and resist harsh environments further restrains the dissolution of the parent substance and therefore further limits the bioavailability of the metals contained in the substance (Environment Canada 2007a).

The total dissolved metal concentrations in experimental solutions assumed to be saturated are reported to vary from 0.012 to 0.179 mg/L for chromium and from 0.02 to 0.36 mg/L for lead (Table 2). The total dissolved metal concentrations were obtained based upon the following procedure: (1) dissolution of the parent substance C.I. Pigment Yellow 34 after 20 to 24 hours stirring in dilution test water of pH 7.1 to 8.4 at room temperature; (2) 0.2 or 0.45 µm filtration and; (3) measurement of total dissolved metals in filtrate. The loading rate was 100 or 125 mg of parent substance per litre (Study Submission 2006a; 2006b; 2006c).

Partitioning

As a metal-containing inorganic substance, a fate analysis based on log K_{ow} and K_{oc} is not applicable to C.I. Pigment Yellow 34. Typical fugacity modelling is also not applicable to this substance or to the metal ions that are released from it on dissolution because, as for other non-volatile chemicals, these substances exert zero partial pressure and fugacity in air (Diamond et al. 1992). The fate of the dissociated metals, however, may be characterized by partition coefficients - namely soil, suspended particle and sediment to water partition coefficients (K_{sw} , K_{ssw} and K_{sdw}), which are presented in Table 2.

Since C.I. Pigment Yellow 34 is a solid and has a negligible vapour pressure, it is not expected to partition to air. Once released to surface water and moist soils, the fate of this substance depends upon its solubility and dissociation in water (OECD 2001) as mentioned above. The fate analysis of the dissociated Pb²⁺ and CrO₄²⁻ ions indicates that they can transform and form dissolved complexes with dissolved ligands present in the aquatic environment (Tipping 2002; Schecher and MacAvoy 1992). Information on metal binding to dissolved ligands may be obtained from thermodynamic constant databases (Smith and Martell 2004; IUPAC 2001), assuming that the metal ion and ligand in the environment are at equilibrium (Stumm and Morgan 1996). Because of the strong tendency of these metals to sorb solid particles in aquatic media (Table 2; K_{ssw}), a significant proportion of dissolved forms of these metals will end up in sediments (Table 2; K_{sdw}), through the settling of suspended particles (Hamilton-Taylor et al. 1984). The remaining metal ions can then be taken up by aquatic organisms as assumed by models relating metal concentrations in aquatic organisms to those of their surroundings (e.g., the free ion activity model, Campbell 1995; the biotic ligand model, DiToro et al. 2001, 2005). Therefore, the moieties of concern issued from the dissolution and dissociation of C.I. Pigment Yellow 34, Pb²⁺ and CrO₄²⁻, are expected to be found in water, sediments and soils but not in air. Note that some nondissolved C.I. Pigment Yellow 34 (as the parent substance) is also expected to be found in sediments and moist soils. When released to dry soils, C.I. Pigment Yellow 34 will mainly remain there with some of the substance leaching locally into ground and/or surface water ecosystems when the soil gets soaked by rain or melting snow/ice. The solid parent substance is not expected to be found in significant amounts in water, considering that its density is a few times greater than that of water.

According to the mass flow tool results presented in Table 5, C.I. Pigment Yellow 34 is mainly released to water during manufacture, formulation, industrial use and service life (35.3%); therefore, releases to water seem to be of greatest potential concern in Canada. As noted above, once released to water, a significant portion of the substance will end up in sediments. Particles of traffic striping paint containing the substance may enter surface water from surface runoff from roadways into drainage ditches, watercourses or wetlands. Additionally, a very small percentage of the substance may be solubilized to release the Pb²⁺ and CrO₄²⁻ moieties of concern. If wastewaters are treated prior to release, a certain percentage of C.I. Pigment Yellow 34 may be removed during the sewage treatment process and later applied to soil as sludge, resulting in release to soil. Of the 62.7% of C.I. Pigment Yellow 34 that is sent to waste disposal (buried in landfills), most will remain in the soil of

the landfill but some leaching (e.g., associated with the degradation of plastics, paints or coatings) might be possible. Overall, considering the information on releases and fate, water and sediments are expected to be the most important receiving media for this chemical.

Data on environmental concentrations of C.I. Pigment Yellow 34 have not been identified. However, concentrations of lead and chromium have been measured in the Canadian environment as natural background and as contamination (e.g., in surface waters, Borgmann et al. 2007).

Persistence and Bioaccumulation Potential

Environmental Persistence

A metal ion is considered infinitely persistent because it cannot degrade any further. When a metal ion is the moiety of concern, the persistence of this metal ion is attributed to the parent compound. This method is justified because even sparingly soluble compounds release a small quantity of metal ion into solution (Environment Canada 2003).

The substance C.I. Pigment Yellow 34 is considered persistent because both of its moieties of concern, the lead (Pb²⁺) and the chromate (CrO₄²⁻) ions are considered infinitely persistent. Therefore, C.I. Pigment Yellow 34 meets the persistence criteria as set out in the *Persistence and Bioaccumulation Regulations* (Canada 2000).

Potential for Bioaccumulation

The current state of the science does not allow for the unambiguous interpretation of the significance of various measures of bioaccumulation (e.g., BCFs, BAFs) for metal-containing inorganic substances. Therefore, such substances are evaluated only on the basis of their properties relating to toxicity and persistence (Environment Canada 2003). It is anticipated that evolution of scientific understanding will eventually allow broader interpretation of the potential for bioaccumulation for such substances.

Potential to Cause Ecological Harm

Ecological Effects Assessment

A - In the Aquatic Compartment

Aquatic acute toxicity of C.I. Pigment Yellow 34 was determined using an alga, *Scenedesmus subspicatus*, an invertebrate named *Daphnia magna* and Rainbow Trout, *Oncorhynchus mykiss*. Table 6 shows the results of these tests.

Table 6. Empirical data	for aquatic toxicit	y of C.I. Pigment Yellow 34

Test organism	Type of test	Endpoint	Duration (hours)	Value (mg/L)	Reference
Scenedesmus subspicatus (Algae)	Acute	EC ₅₀ ¹	72	> 100 ²	Study Submission 2006a
Daphnia magna (Invertebrate)	Acute	EC ₅₀ ³	48	> 100 ²	Study Submission 2006b
Oncorhynchus mykiss (Rainbow Trout – Fish)	Acute	LC ₅₀	96	> 100 ²	Study Submission 2006c

¹Effect on growth rate or development of biomass.

Green algae, *Scenedesmus subspicatus* cultures, were exposed to C.I. Pigment Yellow 34 (Study Submission 2006a). The tests were run at a water temperature of 23 °C and a pH of 7.1 on average. The water hardness level was not mentioned. The substance C.I. Pigment Yellow 34 was added to deionized water, stirred during 20 hours and the undissolved test substance was removed by filtration (0.2 μm). Total dissolved metals (Cr, Pb) were measured in the filtrate. The highest nominal test concentration was 100 mg/L and total dissolved metal concentrations are shown in Table 2. Organisms were exposed to the full strength filtrate (100 mg/L nominal) and subsequent dilutions of the filtrate to give nominal concentrations of 50, 25, 12.5, 6.25, 2.08, 0.69, 0.23 and 0.08 mg/L. There were no significant effects on the reduction in biomass or on the growth rate of algae in any of the concentrations tested after 72 hours.

The substance C.I. Pigment Yellow 34 was also tested on *Daphnia magna* (Study Submission 2006b). The water temperature during the test varied between 19.5 and 20.4°C, pH oscillated between 7.4 and 8.1 and the hardness was in between 220 and 241 mg/L as CaCO₃. The test substance was added to test medium, stirred during 24 hours and the undissolved test substance was removed by filtration (0.2 µm). Total dissolved metals (Cr, Pb) were measured in the filtrate. The highest nominal test concentration was 100 mg/L and total dissolved metal concentrations are shown in Table 2. Organisms were exposed to the full-strength filtrate (100 mg/L nominal) and subsequent dilutions of the filtrate to give nominal concentrations of 50, 25 and 12.5 mg/L. No effects were observed: all daphnids were mobile after 48 hours in each of the concentrations tested.

Finally, the pigment was tested on fish. Juvenile Rainbow Trout, *Oncorhynchus mykiss* were exposed during 96 hours to a moderately hard groundwater containing the test substance (Study Submission 2006c). Average hardness and pH of the test water were 230 mg/L as $CaCO_3$ and 8.3, respectively. The water temperature ranged from 13 to $17^{\circ}C$. Each of the test concentrations (1000; 100; 10 and 1 mg/L) were prepared as follows. All test solutions containing the various amounts of test substance were stirred overnight, followed by 2 filtrations (1 and 0.45 μ m) to remove the undissolved test substance. Average total dissolved metals (Cr, Pb) were measured in the filtrate of the nominal 100 mg/L test concentration. The results are presented in Table 2. Fish were exposed to the filtrates and no mortality was observed in any of the concentrations tested after 96 hours.

²No effect at 100 mg/L loading rate, the highest nominal concentration tested.

³Immobilization.

All laboratory studies showed that there were no effects observed on aquatic organisms at measured concentrations resulting from a 100-mg/L loading rate of the parent substance. It should be noted that the proportion that dissolves could be higher under different environmental conditions. However, because 100 mg/L is a much higher loading rate than would be expected to occur in nature, the concentrations tested can be considered to represent a "reasonable worst-case scenario." Total dissolved metal concentrations as high as those measured in the studies are unlikely to result very often in nature from dissolution of C.I. Pigment Yellow 34. C.I. Pigment Yellow 34 is therefore considered not to be highly hazardous to aquatic organisms.

B - In Other Environmental Compartments

No empirical or predicted effects data for non-aquatic non-mammalian organisms were considered for the metal moieties of concern (lead and chromium) released from the substance. Given the low solubility of the substance, however, exposures for such organisms through contaminated soils and/or food, are not likely to be significant. Laboratory studies on mammals are discussed under the "Potential to Cause Harm to Human Health" section in this screening assessment.

Characterization of Ecological Risk

Manufacture and importation volumes of C.I. Pigment Yellow 34 into Canada along with its widespread and some specific potentially dispersive uses indicate a potential for release of this chemical into the Canadian environment. Based on the available information relating to the metal moieties released on dissolution/dissociation, C.I. Pigment Yellow 34 is persistent in the environment. However, once in the environment, release of bioavailable metals of concern (lead and chromium) will be limited because of its resistance to solubilisation.

Although quantitative estimates of metal exposure resulting from release of C.I. Pigment Yellow 34 have not been made, the experimental ecotoxicological aquatic data suggest that it is not likely to cause acute harm to aquatic organisms, due to its very low water solubility. Although it is possible that dissolution of C.I. Pigment Yellow 34 could result in dissolved concentrations of lead and chromium capable of causing chronic harm to sensitive aquatic biota, the likelihood of this happening is considered to be small. Information on potential impacts in other environmental compartments has not been evaluated; however, considering its low solubility, it is unlikely that organisms associated with other compartments would be harmed by exposure to this substance.

This information suggests that C.I. Pigment Yellow 34 has a relatively low potential to cause ecological harm in Canada.

Uncertainties in Evaluation of Ecological Risk

There is uncertainty regarding metal exposures that could result from dissolution of C.I. Pigment Yellow 34 released into the Canadian environment. Solubility data were experimentally determined after relatively short periods of dissolution time and with neutral to slightly alkaline test water pHs. Longer periods and/or lower pHs representing certain environmental conditions might have led to increased total dissolved metal concentrations in the test solutions. However, using two loading rates (1000 and 100 mg/L) gave similar total dissolved metals concentrations in the test solutions, suggesting that solutions were near or saturated at a pH of around 8.3 (Study Submission 2006c). Solutions were assumed saturated in the experimental conditions tested.

There is also uncertainty about the acute toxicity values that were determined in water with relatively high ionic strength (e.g., high hardness) that could have mitigated metal toxicity (Study Submission 2006b; 2006c). Differences in water chemistries (e.g., hardness, DOC, major ions concentrations, etc.) could then have interfered with bioavailability of the pigment and consequently the potential for adverse effects. On the other hand, some fine particles might have passed through the filters and this might have resulted in increased metal concentrations in the filtrate. This could have resulted in an overestimate of the chemical solubility and bioavailability of the parent substance. Indeed, the particles size distribution presented in Table 2 indicates that a significant proportion of the particles of the analogue substance are smaller than $0.45~\mu m$ and would represent the dissolved colloidal fraction.

Potential for toxicity to soil and sediment-dwelling biota was not specifically examined. However, it is assumed that the conclusion that effects to aquatic species are unlikely because of the limited solubility of the substance (leading to low bioavailable metal fractions) is applicable to other types of organisms as well.

Furthermore, no chronic toxicity tests were conducted for C.I. Pigment Yellow 34 and concentrations of both lead and chromium reported in Table 2 are in the range that could cause chronic effects to sensitive pelagic biota. However, concentrations as high as those measured would be unlikely to result very often in nature from dissolution of C.I. Pigment Yellow 34.

Finally, there is uncertainty in concluding outside a moiety-based assessment on individual substances that can contribute to the total release of bioavailable lead or chromium to the environment. In the context of this Challenge assessment, only sources of bioavailable lead and chromium attributable to dissolution of C.I. Pigment Yellow 34 were considered independently of other sources of these metals to the environment. The conclusion reached in this assessment, that C.I. Pigment Yellow 34 has a relatively low potential to cause ecological harm in Canada, does not preclude the possible inclusion of this substance in future moiety-based assessments of lead or chromium-containing compounds.

Potential to Cause Harm to Human Health

Exposure Assessment

Measured concentrations of C.I. Pigment Yellow 34 in environmental media in Canada or elsewhere were not available. Submissions from a survey conducted under Section 71 of CEPA 1999, in 2006 indicated that industrial releases of C.I. Pigment Yellow 34 to ambient air, water and soil were extremely low (i.e., less than 0.1% of the total quantity manufactured in or imported into Canada) (Environment Canada 2007b). However, the mass flow tool suggests that application and/or post-application releases of the substance may be significant.

Given its physical and chemical properties, estimations of the concentration of C.I. Pigment Yellow 34 in drinking water were not considered to be relevant. Its low solubility in water indicates that the majority of releases to this medium, from industrial or post-application commercial sources (e.g., deterioration of traffic striping paint and migration from landfills after waste disposal), would be in the form of suspended particulates. These particulates are highly likely to be removed via settling or filtration during wastewater and/or subsequent drinking water treatment processes, and therefore the concentrations remaining in drinking water would be negligible.

Due to its negligible vapour pressure, any industrial releases of C.I. Pigment Yellow 34 to ambient air would be in the form of particulates. The majority of these particulates would be captured prior to release from the facility and any remaining particulates would be expected to settle before significant transport had occurred. Using the quantity of releases reported under section 71 of CEPA 1999, a dispersion model was applied using extremely conservative assumptions to estimate the ambient air concentrations downwind from pigment manufacturing facilities (the details of the model cannot be revealed due to the confidential nature of the release data used for the calculations). The maximum concentration of C.I. Pigment Yellow 34 predicted to be present in ambient air was $0.66~\mu g/m^3$. This concentration is expected to be extremely conservative and would only be relevant for those living next to the facility. The general population of Canada is not expected to be exposed to C.I. Pigment Yellow 34 through inhalation of ambient air.

Given the physical chemical properties of C.I. Pigment Yellow 34, in the environment it is expected to be found primarily in soils and/or sediments. Reported industrial releases of this substance to the environment via manufacturing or formulation were extremely low and so are not expected to contribute to the overall concentrations found in soil. As the mass flow tool suggests, application and/or post-application releases (i.e., painting of buildings, vehicles, signage and traffic striping) are likely to be a significant source of this substance to water and eventually to sediments. However, given that the substance is often directly incorporated into the matrix of the solid material and the dispersive use of these products, the availability of this substance for human exposure is expected to be minimal. Landfills and areas of high product usage may have higher concentrations of this substance in the soil; however, migration of the substance from these areas is expected to be low due to its negligible vapour pressure and low water solubility. The low solubility of this substance also

indicates that bioavailable exposures through soils, suspended solids and sediment are not likely to be significant and would be further reduced by the substance being incorporated into a solid matrix or encapsulated in silica. Therefore significant quantities of C.I. Pigment Yellow 34 are not expected to be found in the food chain.

The substance C.I. Pigment Yellow 34 is not used for the manufacture of products for use by the general population and therefore there is no direct exposure to them via this route. This substance is used in commercial settings to manufacture items that consumers may come in contact with after application, such as road paint, pigmented polymers and pigments used in wiring. As the substance is not volatile, there would be no relevant exposure through the inhalation pathway. It is possible that a consumer may have dermal contact with the pigments following application; however, the resulting dermal exposure is expected to be low for several reasons. This substance is often directly incorporated into the matrix of the solid material (i.e., polymer) and, generally, solid materials have the lowest potential for exposure by the dermal route as migration through the solid matrix and subsequent absorption through the skin would be very limited. Specifically, chromium and lead, and particularly their salts, are not known to have a high potential for systemic exposure by the dermal route as they have low skin permeability coefficients relative to other chemicals (US EPA 1992) and the silica encapsulation of this pigment would further prevent migration.

The confidence in the characterization of exposure to C.I. Pigment Yellow 34 is low as the concentrations of this substance in environmental media could not be quantified in Canada or elsewhere. However, given the physical and chemical properties of this substance and its commercial use and applications, it is expected that exposure to the general population of Canada is negligible. It is possible that there may be small populations of Canadians that are exposed to higher concentrations than would be expected in the general environment. This could occur in persons living near facilities that are manufacturing or using large quantities of this substance or those that are participating in activities such as automotive restoration and repair, or using artists' paints that may contain C.I. Pigment Yellow 34. Given the lack of data available for this substance, it is not possible to quantify the exposures from these sources.

Health Effects Assessment

The available experimental toxicological data on health effects for C.I. Pigment Yellow 34 and its principal components, lead chromate and lead sulfate, as well as the epidemiological investigations among workers involved in lead chromate pigment production and use are presented in Appendix 1.

The European Commission (ESIS 2007) has classified C.I. Pigment Yellow 34 as a Category 3 carcinogen ("causes concerns for humans owing to possible carcinogenic effects")¹. The European Commission has concluded that C.I. Pigment Yellow 34, together with lead chromate and C.I. Pigment Red 104, "show(s) evidence for carcinogenicity in several studies

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¹ The European Commission has proposed a reclassification for C.I. Pigment Yellow 34 as a Category 2 carcinogen ("should be regarded as if they are carcinogenic") (ECB 2007).

with rats after subcutaneous and intramuscular administration. Lead chromate induced both benign and malignant tumours at the site of injection and, in one study, renal carcinomas. The animal studies are supported by epidemiological studies demonstrating an increased frequency of lung cancer among workers involved in production of chromate pigments. The animal studies are also supported by genotoxic(ity) studies as well as cell transformation studies. The substances show resemblance to known mutagens/carcinogens" (ECB 2003). The carcinogenicity of C.I. Pigment Yellow 34 has not been examined in experimental animals via oral, dermal or inhalation administrations. However, bronchial squamous cell carcinomas were observed in male rats in a two-year implantation study, in which various C.I. Pigment Yellow 34 preparations were implanted in the left lung (Levy et al. 1986). Although these results were not statistically significant, the rarity of this lesion in the control animals indicates a certain level of relevance. Additionally, local benign and malignant tumours were significantly induced in male and female rats following subcutaneous or intramuscular injection of lead chromate, the principal component of C.I. Pigment Yellow 34 and in one study 3 of 23 male rats developed renal tumours, which were not observed in control animals (0 of 22) (Maltoni 1974, 1976; Maltoni et al. 1982; Furst et al. 1976). A number of additional studies by intratracheal, intrapleural, intrabronchial and intramuscular administration of lead chromate or its derived pigments did not significantly induce tumour incidence in various animal species (Levy et al. 1986; Hueper 1961; Furst et al. 1976; Steffee and Baetjer 1965).

In humans, epidemiological investigations have been conducted in occupational settings in various geographic locations with an attempt to identify the relationship between occupational engagement in lead chromate pigment production and cancer risk. Workers in this industry were exposed not only to the pigments themselves but also to the soluble hexavalent chromium compounds used as raw materials in the pigment production. The majority of the results showed an increased risk of lung cancer among the workers in the plants where both lead and zinc chromate pigments were produced (Sheffet et al. 1982; Hayes et al. 1989; EEH 1976; EEH 1983; Davies 1979; Davies 1984; Haguenoer et al. 1981; Deschamps et al. 1995; Fentzel-Beyme 1983; Korallus et al. 1993). The only exception is the study conducted in five chromate pigment production plants in Japan where no significantly increased mortality due to lung cancer was observed (Kano et al. 1993). The authors stated this might be because the amount of hexavalent chromium compounds in the work environment has been lowered as a result of engineering hygiene considerations such as improved ventilation, the wearing of masks, attention to work clothes and bathing after work. Two epidemiological studies conduced in the plants where only lead chromate pigments were produced reported a slightly elevated risk in respiratory tract tumour, but no statistical significance has been reached (Davies 1979; EEH 1983). The authors speculated that the numbers of observed and expected deaths were too small in these studies for definitive conclusions. With respect to lead chromate pigment use, the only available epidemiological investigation did not indicate a statistically significant association between spray painting and respiratory-cancer-caused mortality (Chiazze et al. 1980).

In addition to the European Commission classification, lead chromate pigments have been assessed together with hexavalent chromium (chromium VI) compounds by other jurisdictions. The International Agency for Research on Cancer (IARC 1990) classified

chromium VI compounds as Group 1 ("carcinogenic to humans"). IARC concluded that there is "sufficient evidence" in humans for the carcinogenicity of chromium VI compounds as encountered in the chromate pigment production industry and "sufficient evidence" in experimental animals for the carcinogenicity of lead chromates. The United States National Toxicological Program (NTP 2005a) concluded that chromium VI compounds are "known to be human carcinogens based on sufficient evidence of carcinogenicity in humans" and the United States Environmental Protection Agency (US EPA 1998) has classified chromium VI compounds as Group A — "known human carcinogen by the inhalation route of exposure." However, the US EPA concluded that carcinogenicity of chromium VI compounds "by the oral route of exposure cannot be determined" (Group D). Environment Canada and Health Canada (Canada 1994) concluded that "the group of hexavalent chromium compounds as a whole is entering the environment in a quantity or concentration or under conditions that may constitute a danger in Canada to human life or health" and classified chromium VI compounds as "carcinogenic to humans" (Group 1).

Lead chromate has also been assessed together with inorganic lead compounds. The International Agency for Research on Cancer (IARC 2006) classified inorganic lead compounds as "probably carcinogenic to humans" (Group 2A) and concluded that there is "limited evidence" in humans and "sufficient evidence" in experimental animals for the carcinogenicity of inorganic lead compounds. The United States National Toxicological Program (NTP 2005b) concluded that lead compounds are "reasonably anticipated to be human carcinogens based on limited evidence from studies in humans and sufficient evidence from studies in experimental animals" and the US EPA (US EPA 1993) concluded that inorganic lead compounds are "probable human carcinogens" (Group 2B) based on "sufficient animal evidence" while "human evidence is inadequate." Lead is also listed on Schedule 1 (List of Toxic Substances) under the *Canadian Environmental Protection Act*, 1999 in Canada (Environment Canada 2006).

The European Commission has stated that C.I. Pigment Yellow 34, together with C.I. Pigment Red 104 and lead chromate, "show(s) pronounced response in several in vitro assays; one positive in vivo bone marrow test is available. The substances show resemblance to known mutagens/carcinogens" (ECB 2003). The genotoxicity of C.I. Pigment Yellow 34, as well as its principal components, have been assessed in a range of assays (see Appendix 1). In vivo, C.I. Pigment Yellow 34, suspended in olive oil, did not significantly increase the frequency of micronuclei induction in mouse bone marrow cells following intraperitoneal injection; however, the authors speculated that the results may not be definitive as there was no evidence that the test material was reaching the target tissue (Odagiri et al. 1989). Additionally, lead chromate showed positive results in *in vivo* micronuclei induction when suspended in gum Arabic saline, and positive results in sex-linked recessive lethal assay when dissolved in nitrilotriacetic acid (NTA) (Watanabe et al. 1985; Costa et al. 1988). It is recognized that the *in vitro* genotoxicity of C.I. Pigment Yellow 34 and lead chromate are often enhanced following dissolution in acid or alkali or NTA solution. Aqueous suspensions of C.I. Pigment Yellow 34 did not significantly induce gene mutation in bacteria unless dissolved in NaOH or NTA (Venier et al. 1985; De Flora 1981; Connor and Pier 1990; Pier et al. 1991). On the other hand, aqueous C.I. Pigment Yellow 34 (as chromium yellow), showed clastogenic effects in mammalian cells as indicated by chromosomal aberration and

sister chromatid exchange assays (Levis and Majone 1981; Venier et al. 1985), although a recent publication did not indicate that C.I. Pigment Yellow 34 significantly induced chromosomal aberration in CHO cells (Nestmann and Zhang 2007). Lead chromate was also clastogenic in various tests. When suspended in water or acetone, it induced genetic damage in human and animal cells *in vitro*, including chromosomal aberration, micronuclei induction and sister chromatid exchange. It caused chromium-DNA adducts formation, DNA-protein cross-linking, DNA strand breakage, centrosome abnormalities, aneuploidy and cell transformation. Upon primary dissolution, lead chromate induced mutation in bacteria and mammalian cells, and mitotic recombination in yeast (Appendix 1).

Although a thorough analysis of the mode of action is beyond the scope of this screening assessment, the European Commission has stated that a possible mode of action for the genotoxicity of C. I. Pigment Yellow 34 is through the reduction of insoluble PbCrO₄ by glutathione reductase/NADPH and the generation of OH⁻ radicals that cause DNA strand breakage (ECB 2003).

Limited information is available regarding non-cancer endpoints for C.I. Pigment Yellow 34. In the only available studies conducted in experimental animals, C.I. Pigment Yellow 34 preparations, with or without silica-encapsulation, were orally administrated to rats and dogs for 90 days. The lowest-observed-(adverse)-effect level (LO(A)EL) in dogs was observed at 2000 ppm administered in the diet (60 mg/kg bw/day) and was based on significantly altered haematological parameters including decreased haemoglobin and haematocrit values and altered erythrocyte morphology, significantly increased aminolevulinic acid in the urine, as well as dose-related pathological changes of the kidneys. Impaired gonadal development was noted in the exposed animals but not in the controls, which the authors speculated to be the result of poor nutrition or the younger age of the animals at the sacrifice time, and therefore was considered to be an indirect effect of the treatment (IBT 1975a, b, c; IBT, 1976a). Similar haematological, urinary and renal effects were observed in rats administered 2000 ppm in the diet (100 mg/kg-bw/day) (IBT 1975d, e, f; IBT 1976b). These studies were conducted by Industrial Bio-Test laboratories Inc. (IBT), and the abstracts of the study results were published in *Toxicology and Applied Pharmacology* (Christofano et al. 1976; Kennedy et al. 1976). No information is available as to whether these studies have been audited, reducing the confidence of the information presented.

The European Commission (ESIS 2007) has classified C.I. Pigment Yellow 34 as Category1 for developmental toxicity ("known to cause developmental toxicity in humans") and as Category 3 for reproductive toxicity ("causes concern for human fertility"). The original dataset considered for these classifications is not available. A limited epidemiological investigation of a lead chromate production plant in China showed that the rates of threatened abortions were significantly higher among female workers and the wives of male workers who had been exposed to lead chromate (Wang and Zhao 1985). Effects of C.I. Pigment Yellow 34 on the reproductive system were also observed in the oral subchronic studies in dogs, described above, where impaired gonadal development was noted in the treated animals but not in the controls, although the authors considered it as an indirect effect of the treatment (Christofano et al. 1976, original data in IBT 1975a, b, c, 1976a).

The low solubility of C.I. Pigment Yellow 34 is indicative of limited bioavailability. The bioavailability of lead chromate and lead-chromate-derived pigments has been investigated in experimental animals. Administration of non-encapsulated or silica-encapsulated Chrome Yellow/lead chromate to rats by gavage (150 mg/kg-bw/day, five days per week, for four weeks) resulted in an increased level of lead in the blood and kidneys. No chromium could be detected in blood from exposed rats (detection limit = $10 \mu g/L$). The kidney levels of chromium were increased significantly only in rats treated with non-encapsulated pigment. These results indicate that silica encapsulation reduces the gastrointestinal bioavailability of chromium from lead chromate pigments (Clapp et al. 1991; Pier et al. 1991). Administration of lead chromate to rats via whole body inhalation $(5.3 \pm 0.8 \text{ mg CrVI/m}^3, 4 \text{ hours per day})$ for 1 to 4 days) led to the accumulation of both chromium and lead chromate in the lungs. The chromium concentration in urine and feces were significantly increased following administration, whereas both chromium and lead concentrations in blood were only slightly elevated (above 5 µg/L for chromium) (Bragt et al. 1990). In addition, a short-term study in male rats showed that lead did not migrate from polypropylene plastic coloured with lead chromate-molybdate following oral administration (Gage and Litchfield 1967). Investigations employing other routes of administration, including intratracheal injection, instillation and infusion to the tracheal lobe bronchus, of lead chromate or lead paint resulted in increased lead and chromium levels in various tissues and retention in the lungs (Bragt and van Dura 1983; Perrault et al. 1995; Eaton et al. 1984).

Workers in a plastic production plant exposed to dust containing various chemicals, including lead and lead chromate, had significantly increased chromium levels in their urine samples. Their blood lead levels were also significantly increased, but not their serum chromium levels (Boscolo et al. 1997). Other occupational studies (McAughey et al. 1988; Wiegand et al. 1988) also showed that the urine and blood chromium levels in lead chromate pigment production workers were higher than those typically observed in non-occupationally exposed persons (Iyengar and Woittiez 1988). In two lead chromate-based paint factories in the UK, blood lead levels were detected in a range of 9–25 μ g/L for warehouse men, a range of 10–36 μ g/L for ball mill loaders, and a range of 9–15 μ g/L for spray painters. The author stated that these levels were commonly found in non-lead workers (Cowley 1984).

Data from the bioavailability studies in experimental animals and observations in occupationally exposed humans suggest that lead chromate and its derived pigments have some level of bioavailability.

The confidence in the hazard characterization of neoplastic potential associated with C.I. Pigment Yellow 34 is considered to be moderate, as data are available from epidemiological investigations, from carcinogenicity studies in experimental animals via parenteral administration and a number of genotoxicity studies. However, no long-term (> 90 days) laboratory studies that were conducted via conventional administration routes (oral, dermal or inhalation) are available. The confidence in the hazard characterization of non-cancer effects associated with C.I. Pigment Yellow 34 is considered to be low. There is a high level of uncertainty regarding the reproductive toxicity and developmental toxicity of C.I. Pigment Yellow 34 due to lack of information. With respect to the repeated-dose toxicity of C.I. Pigment Yellow 34, characterization of potential toxicological effects is limited as experimental animal data is available for only one route (oral) and for one duration and these

studies were conducted by Industrial Bio-Test Laboratories. In addition, the oral LO(A)EL identified was the lowest exposure level tested in the studies, increasing the uncertainty as to the actual effect level at which these effects may occur.

Characterization of Risk to Human Health

Based principally on the weight of evidence classification of C.I. Pigment Yellow 34 by the European Commission as a Category 3 carcinogen (ESIS 2007), and the assessments of hexavalent chromium and inorganic lead compounds by several national and international agencies (US EPA 1993, 1998; IARC 1990, 2006; Canada, 1994; NTP 2005a, b), a critical effect for characterization of risk to human health for C.I. Pigment Yellow 34 is carcinogenicity. Although most of these classifications were based on groups of hexavalent chromium compounds or inorganic lead compounds, they, along with the information on lead chromate, support the limited data available on C.I. Pigment Yellow 34. As well, C.I. Pigment Yellow 34 or its principal components were genotoxic in a limited number of *in vitro* and *in vivo* experimental systems. Therefore, a mode of action for carcinogenicity involving direct interaction with genetic material cannot be precluded.

With respect to non-cancer effects, margins of exposure comparison were not derived due to negligible potential for exposure to the general population of Canada and the uncertainties regarding non-cancer effect levels (reproductive toxicity, developmental toxicity and repeated-dose toxicity).

Uncertainties in Evaluation of Risk to Human Health

There is uncertainty regarding the bioavailability of this substance; however, limited data indicate that there is some level of absorption of lead and chromate after exposure to lead chromate and its derived pigments. In addition, although genotoxicity of the pigment or lead chromate is generally more pronounced after dissolution in acid or base, positive results were also obtained in aqueous media. However, encapsulation of the pigment has been shown to reduce bioavailability and genotoxicity in some studies. In-depth consideration of the mode of action for carcinogenicity for this substance is beyond the scope of this screening assessment. As well, the health effects database for the C.I. Pigment Yellow 34 as a discrete substance is limited. There is uncertainty concerning to the actual exposure levels of the workers in some of the epidemiological investigations as workplace exposure monitoring data were not available and protective measures were sometimes implemented during the time period of studies.

The confidence in the characterization of exposure to C.I. Pigment Yellow 34 is low as the concentrations of this substance in environmental media could not be quantified in Canada or elsewhere leading to uncertainty in the exposure assessment. However, based on the properties and uses of the substance, there is confidence that actual environmental exposure levels are low. However, for a small population of Canadians, there is uncertainty regarding possible exposure to higher concentrations, but these could not be quantified.

Conclusion

Based on the available information, it is concluded that C.I. Pigment Yellow 34 is not entering the environment in a quantity or concentration or under conditions that have or may have an immediate or long-term harmful effect on the environment or its biological diversity, or that constitute or may constitute a danger to the environment on which life depends.

On the basis of the carcinogenicity of C.I. Pigment Yellow 34, for which there may be a probability of harm at any level of exposure, it is concluded that C.I. Pigment Yellow 34 is a substance that may be entering the environment in a quantity or concentration or under conditions that constitute or may constitute a danger in Canada to human life or health.

It is therefore proposed that C.I. Pigment Yellow 34 does not meet the criteria in paragraph 64a and 64b of CEPA 1999, but it does meet the criteria in paragraph 64c of CEPA 1999. Additionally, C.I. Pigment Yellow 34 meets the criteria for persistence as set out in the *Persistence and Bioaccumulation Regulations* (Canada 2000).

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Appendix 1. Summary of health effects information for C.I. Pigment Yellow 34 (CAS RN: 1344-37-2)¹

Endpoint	Lowest effect levels/Results ²		
CASRN	1344-37-2 (C.I. Pigment Yellow 34; Chrome orange)	7758-97-6 (lead chromate)	7446-14-2 (lead sulfate)
Laboratory anima	als and <i>in vitro</i> ³		
Acute toxicity	Lowest oral LD ₅₀ (rat) > 5000 mg/kg-bw (NPIRI <i>Raw Materials Data Handbook</i> 1983).	Lowest oral LD50 (mouse) > 12 000 mg/kg-bw (Yakuri 1968).	Lowest oral LD _{LO} (dog) = 2000 mg/kg-bw (equivalent to 1366 mg lead/kg-bw, Sax
	[Additional study in rats, BASF AG 1974]	[No additional study identified]	1984).
	[The skin and eye irritation tests in rabbits were negative (Ciba-Geigy Limited 1983)]		[Additional study in guinea pigs, Sax 1984]
Short-term repeated-dose toxicity	No data identified.	No data identified.	No data identified.
Subchronic toxicity	Lowest oral LOEL (dog, 4/sex/group, 90 days) = 2000 ppm in diet, equivalent to 60 mg/kg-bw/day, based on significantly decreased haemoglobin and haematocrit values, altered erythrocyte morphology, pathologic changes of the kidney. Other effects and clinical signs such as impaired gonadal development, lethargy, anorexia, dehydration, emaciation, hyperirritability, disorientation, motor ataxia and convulsions prior to death were observed (Christofano et al. 1976, original data in IBT 1975a, b, c, 1976a). [Additional studies in rats: Kennedy et al., 1976, original data in IBT 1975d, e, f, 1976b]	No data identified.	No data identified.
Chronic toxicity/ Carcinogenicity	Carcinogenicity: Bronchial implantation of 2 mg of primrose chrome yellow, light chrome yellow, LD chrome yellow supra 70FS, medium chrome yellow and medium chrome yellow-silica encapsulated, respectively, in left lung of Porton-Wistar rats (50/sex/groups). Animals were maintained for 2 years. Bronchial carcinomas developed in one male rat in each of the primrose chrome yellow, light chrome yellow and medium chrome yellow groups.	Carcinogenicity: Positive results Single subcutaneous injection of 30 mg lead chromate or basic lead chromate to Sprague-Dawley rats (20/sex/group). Animals were maintained for 117 to 152 weeks. Significant increases of sarcomas at the injection sites were observed (26/40 or 27/40 in treated animals versus 0/60 or 1/80 in control animals) (Maltoni	No data identified.

Endpoint	Lowest effect levels/Results ²			
CASRN	1344-37-2 (C.I. Pigment Yellow 34; Chrome orange)	7758-97-6 (lead chromate)	7446-14-2 (lead sulfate)	
CASRN	The result was not statistically significant. Bronchial inflammation, squamous metaplasia and dysplasia were reported (Levy et al. 1986). [No additional study identified]	Intramuscular injection of 8 mg lead chromate to Fisher-344 rats (25/sex/group) once a month for 9 months resulted in significant increases of local sarcomas (31/47 in treated animals versus 0/22 in control animals). In addition, renal carcinomas were observed in male rats at 24 months (3/23 in treated animals versus 0/22 in control animals) (Furst et al. 1976). Negative results After intramuscular injection of lead chromate (doses unspecified) to rats (32–34/group), 1/33 rats developed local tumours. The result was not statistically significant. No further information available (Hueper 1961). After intrapleural implantation of lead chromate (doses unspecified) in rats for 27 months, 3/34 rats developed local tumours. The result was not statistically significant (Hueper 1961). After intramuscular injection of 3 mg lead chromate to NIH-Swiss mice (25 female/group) every four months for two years, lymphonmas were observed at 16 months (2/17 in treated animals versus 1/15 in untreated control and 2/22 in vehicle treated control), lung carcinomas were observed at 24 months (3/17 in treated animals versus 1/15 in untreated control and 1/22 in vehicle-treated control). The result was not statistically significant (Furst et al. 1976).	7446-14-2 (lead sulfate)	
		Intra-tracheal instillation of 3 mg lead chromate		

Endpoint	Lowest effect levels/Results ²		
CASRN	1344-37-2 (C.I. Pigment Yellow 34; Chrome orange)	7758-97-6 (lead chromate)	7446-14-2 (lead sulfate)
		preparation in guinea pigs (13/group) six times at 3-month intervals. The animals were maintained for life. No pulmonary adenoma was observed (Steffee and Baetjer 1965).	
		Intra-tracheal instillation of 10-mg lead chromate preparation in rabbits (7/group) every 3 months for 9–15 months. The animals were maintained for 40–50 months. No pulmonary tumours observed (Steffee and Baetjer 1965).	
		Bronchial implantation of 2 mg lead chromate in Porton-Wistar rats (50/sex/groups). The animals were maintained for two years. Bronchial tumours were observed (1/98 in treated animals versus 0/100 in control animals). The result was not statistically significant (Levy et al. 1986).	
Reproductive toxicity	No data identified.	No data identified.	No data identified.
Developmental toxicity	No data identified.	No data identified.	No data identified.
Genotoxicity and related endpoints: in vitro	Mutagenicity Positive in Ames tests in Salmonella typhimurium TA 100 when dissolved in nitrilotriacetic acid or NaOH, with or without metabolic activation. Negative in Salmonella typhimurium TA98, TA100, TA1535, TA1537 and TA 1538, with and without metabolic activation. Test material was dissolved in ether or encapsulated with high percentages of silica (for TA100) (De Flora et al. 1981; Connor and Pier 1990; Pier et al. 1991). Sister Chromatid Exchange Positive in CHO cells when suspended in water or dissolved in NaOH or nitrilotriacetic acid (NTA) (Levis	Mutagenicity Ames tests Positive in Salmonella typhimurium TA 98 and TA1538, with and without metabolic activation; Positive in TA 100, with metabolic activation; Positive in TA1537, without metabolic activation. Test material was dissolved in HCl (Nestmann et al. 1979). Negative in Salmonella typhmurium TA1535, with and without metabolic activation; Negative in TA1537, with metabolic activation or encapsulated with high percentages of silica;	Mutagenicity Negative: Ames tests in Salmonella typhimurium TA98, 100, 1535, 1537, 1538 with and without activation (JETOC 1996; Connor and Pier 1990). Mutation in E. coli wp2uvra with and without activation (JETOC 1996).
	dissolved in NaOH or nitrilotriacetic acid (NTA) (Levis and Majone 1981; Venier et al. 1985).	Negative in TA102, without metabolic activation. Test material was suspended in water or oil, or	Sister Chromatid Exchange Positive:

Endpoint	Lowest effect levels/Results ²		
CASRN	1344-37-2 (C.I. Pigment Yellow 34; Chrome orange)	7758-97-6 (lead chromate)	7446-14-2 (lead sulfate)
	Chromosomal aberration Positive in CHO cells. Test material (chromium yellow) was suspended in cell growth medium or dissolved in NaOH (Levis and Majone 1981); Negative in CHO cells. A small increase in aberrations was observed but the result was not statistically significant. C.I. Pigment Yellow 34 was suspended in water. Dose-related cytotoxicity and mitotic inhibition was observed (Nestmann and Zhang 2007).	dissolved in HCl or DMSO (Nestmann et al. 1979; De Flora et al. 1985; Pier et al. 1991). Gene mutation in Escherichia coli Positive in E. coli K12 WP2 uvr trp ⁺ reversion fluctuation assay, without metabolic activation. Test material was dissolved in 0.5 N NaOH (Nestmann et al. 1979); Positive in E. coli trp ⁻ to trp ⁺ spot tests and fluctuation test when dissolved in nitrilotriacetic acid, but Negative when suspended in water (Venier et al. 1987, cited in ECB 2003); Negative in E. coli K12 gal ⁻ forward mutation and E. coli trp ⁻ reversion plate test, without metabolic activation, when dissolved in 0.5 N NaOH (Nestmann et al. 1979). Gene mutation in mammalian cells Positive in HPRT mutation assays in CHO-K1 cells when dissolved in 1N NaOH (Yang et al. 1992, cited in ECB 2003); Negative in HPRT mutation assays in Chinese hamster V79 cells for 8-azagunine resistance and 6-thioguanine resistance when suspended in water or cell culture medium but Positive results were obtained when dissolved in NTA (Newbold et al. 1979 and Celotti et al. 1987); Negative in mutation assays in C3H 10T1/2 mouse cells for ouabain resistance, and in CHO cells for 6-thioguanine resistance and ouabain resistance, without metabolic activation, test material was suspended in acetone (Patierno et al. 1988; Patierno and Landolph 1989). Chromosomal Aberration Positive in CHO cells and human foreskin	In human lymphocytes cells and in CHO cells, test material was suspended in water or mixed with NTA (Tucker et al. 1993; Montaldi et al. 1985).

Endpoint	Lowest effect levels/Results ²		
CASRN	1344-37-2 (C.I. Pigment Yellow 34; Chrome orange)	7758-97-6 (lead chromate)	7446-14-2 (lead sulfate)
		fibroblasts, without metabolic activation (Xu et al. 1992; Wise et al. 1992, 1994, cited in ECB 2003 and Grlickova-Duzevik et al. 2006; Savery et al. 2007); Positive in human primary bronchial fibroblasts (Wise et al. 2002), human lung cell line WTHBF-6 cells and human epithelial cell line BEP2D cells (Wise et al. 2004, 2006), the test material was suspended in acetone in all the experiments; Positive in CHO cells when the test material was suspended in water (Koshi and Iwasaki 1983) or dissolved in NTA (Montaldi et al. 1987) and in human lymphocytes when the test	
		material was dissolved in NaOH (Douglas et al. 1980) Micronuclei induction Positive in human peripheral blood lymphocytes. An enhanced effect was observed following addition of NTA (Montaldi et al. 1987).	
		Sister Chromatid Exchange Positive in human peripherical blood lymphocytes when suspended in water or dissolved in NaOH (Tucker et al. 1993, cited in ECB 2003; Douglas et al. 1980). Positive in CHO cells when suspended in water or dissolved in NaOH or NTA (Loprieno et al. 1985; Montaldi et al. 1987).	
		DNA damage Positive: DNA single strand breaks or DNA-protein crosslinks in CHO cells, without metabolic activation. Test material was suspended in acetone (Xu et al. 1992).	

Endpoint Lowest effect levels/Results ²			
CASRN	1344-37-2 (C.I. Pigment Yellow 34; Chrome orange)	7758-97-6 (lead chromate)	7446-14-2 (lead sulfate)
		Comet assay for double strand DNA breaks in human bronchial cells. Test material was suspended in acetone (Xie et al. 2005).	
		Apoptosis and DNA fragmentation in CHO cells. Test material was suspended in acetone (Blankenship et al. 1997).	
		Negative: DNA double strand breaks or DNA-DNA crosslinks. Test material was suspended in acetone (Xu et al. 1992).	
		Differential survival tests in <i>E. coli</i> W3110 (<i>polA</i> ⁺) and P3478 (<i>polA</i> ⁻). Test material was dissolved in 0.5 N NaOH (Nestmann et al. 1979).	
		Induction of DNA fragmentation in CHO cells. Test material was dissolved in NaOH (Douglas et al. 1980).	
		Cr-DNA adduct formation Positive in human lung small airway epithelial cells (Singh et al. 1999).	
		SOS response Negative in <i>E. coli</i> PQ37 when dissolved in water but Positive when dissolved in NTA (Vernier et al. 1989).	
		Cell transformation Positive in the C3H 10T1/2 mouse cells and Syrian hamster embryo cells, with and without simian adenovirus SA7 viral enhancement, and	

Endpoint	Lowest effect levels/Results ²			
CASRN	1344-37-2 (C.I. Pigment Yellow 34; Chrome orange)	7758-97-6 (lead chromate)	7446-14-2 (lead sulfate)	
		without metabolic activation; Positive in human non-tumorigenic osteosarcoma TE85 cells, without metabolic activation. Test material was suspended in acetone or water or cell culture medium (Patierno et al. 1988; Patierno and Landolph 1989; Schechtman et al. 1986; Elias et al. 1989; Sidhu et al. 1991); Positive in induction of anchorage independent colonies formation in human foreskin cells. Test material was		
		suspended in acetone (Biedermann and Landolph 1987). Mitotic recombination Positive in Saccharomyces cerevisiae D5 mitotic recombination, without metabolic activation. However, S9 reduced the mutagenic potency. Test material was dissolved in 1 N HCl (Nestmann et al. 1979).		
		Centrosome abnormalities and aneuploidy Positive in human WTHBF-6 cells (a clonal cell line from normal human bronchial fibroblasts) when suspended in acetone (Holmes et al. 2006; Wise et al. 2006)		
Genotoxicity and related endpoints: in vivo	Micronuclei induction: Negative in ICR/Jcl mouse bone marrow cells following intraperiotoneal injection with chrome-yellow. Test material was suspended in olive oil. The authors speculated that the results may not be definitive as there	SLRLM (sex-linked recessive lethal mutation) Positive in <i>Drosophila melanogaster</i> . Test material was dissolved in NTA (Costa et al. 1988) Micronuclei induction	No data identified.	
	was no evidence that the test material was reaching the target tissue (Odagiri et al. 1989). [No additional study identified]	Positive in C57B1/6N mouse bone marrow cells following intra-peritoneal injection. Test material was suspended in 0.5% gum Arabic saline (Watanabe et al. 1985).		

Endpoint	Lowest effect levels/Results ²		
CASRN	1344-37-2 (C.I. Pigment Yellow 34; Chrome orange)	7758-97-6 (lead chromate)	7446-14-2 (lead sulfate)
Chronic toxicity/	Neoplastic endpoints	Neoplastic endpoints	Neoplastic endpoints
carcinogenicity	Lead chromate pigment production	See data for 1344-37-2	See data for 1344-37-2
	Historical cohort study among men employed at a lead		
	and zinc chromate pigment production plant (lead to zinc		
	= 9:1) in the United States during 1940–1969 for more		
	than 1 month. 1296 whites and 650 non-whites were		
	included in the study. A significantly increased relative		
	risk (RR) for lung cancer (RR = 160) was observed for		
	the white men. The RR was increased to 190 for the		
	white men who were employed for at least 2 years and		
	who had exposed to 0.5 to > 2 mg of CrVI/m ³ chromates.		
	Although RR of 200 for stomach cancer, 170 for		
	pancreatic cancer, and 290 for Hodgkin's disease were		
	found, none of these were significant. However, further		
	analysis revealed a significant risk for stomach cancer in		
	the white men [standardized mortality ratio (SMR) =		
	[230] and lung cancer in the non-white men (SMR = 200).		
	Air monitoring at the plant in the later years were from		
	less than 0.1 to more than 2 mg of CrVI/m ³ . Workers		
	were probably co-exposed to nickel sulfate and nickel		
	carbonate. Smoking information was not available. 67		
	subjects were removed in the follow-up study conducted		
	in 1982. The increased risk for lung cancer was observed		
	to be associated with the years of employment and the		
	years of exposure to chromate dusts in the factory. For		
	the subjects who had been followed for more than 30		
	years and had been employed for more than ten years, the		
	SMR for lung cancer was 190 (95% $CI = 111-295$), and		
	among this group, for those whose jobs were associated		
	with exposure to chromate dust, the SMR was increased		
	to 321 (95% CI = 117-698) (Sheffet et al. 1982; Hayes et		
	al. 1989).		
	Historical ashort study among 574 man amplement at		
	Historical cohort study among 574 men employed at		
	three chromate pigment production plants in the United		

Endpoint	Lowest effect levels/Results ²		
CASRN	1344-37-2 (C.I. Pigment Yellow 34; Chrome orange)	7758-97-6 (lead chromate)	7446-14-2 (lead sulfate)
	States from the mid-1920s to the end of 1979 for at least		
	6 months. At plant 1 (246 workers) where lead chromate		
	pigments (chrome yellow, chrome green and molybdata		
	orange) were produced, the SMR for respiratory cancer		
	was $164.4 (95\% CI = 45-421)$; for lung cancer, it was 130		
	(95% CI = 27-381); for digestive system cancer, it was		
	120.3 (95% CI = 15-434); for all causes, it was 63.5		
	(95% CI = 40-97). At plant 2 (164 workers) and 3 (164		
	workers), both lead and zinc chromate pigments were		
	produced. In addition, at plant 2, strontium chromate and		
	barium chromate were produced. The workers in plant 3		
	were also studied by Sheffet et al. 1982 (see above). The		
	combined SMR from plants 2 and 3 for lung cancer or for		
	stomach cancer was significantly increased (SMR = 228		
	and 667, respectively). Because of the non-significant		
	rate for respiratory cancer at plant 1 and the co-exposure		
	to other chromates at plants 2 and 3, no definitive		
	conclusion regarding the risk of lung cancer in lead		
	chromate-exposed workers can be drawn from this study.		
	In 1975, the average workroom air concentrations of		
	chromium and lead in plant 1 were 0.05 mg total		
	chromium/m ³ and 0.28 mg lead/m ³ ; in plant 2, 0.06 mg		
	total chromium/m ³ and 0.26 mg lead/m ³ ; in plant 3, 0.19		
	mg chromium/m ³ and 0.79 mg lead/m ³ [EEH 1976; EEH		
	1983].		
	Historical cohort study among 1152 men who worked at		
	three chromate pigment production plants in the United		
	Kingdom for at least 1 year from the 1930s or 1940s to		
	1981. Significant excesses for lung cancer mortality were		
	observed only among the workers exposed to high and		
	moderate levels of chromates at factories A (675 men)		
	and B (222 men) where zinc and lead chromate pigments		
	were produced at both factories. The SMR for lung		
	cancer was significantly increased in both factories (SMR		

Endpoint	Lowest effect levels/Results ²		
CASRN	1344-37-2 (C.I. Pigment Yellow 34; Chrome orange)	7758-97-6 (lead chromate)	7446-14-2 (lead sulfate)
	was in the range of 222 to 223 in factory A; and 373 to		
	562 in factory B). The combined SMR for lung cancer in		
	the medium and high exposure groups was 310 (95% CI		
	= 160-540), 270 (95% CI = 110-560), or 388 (95% CI =		
	170-760) for 1 year, 5 to 9 years or more than 10 years		
	employment, respectively. 255 men at factory C were		
	exposed only to lead chromate and experienced normal		
	mortality. No exposure monitoring data of chromate or		
	lead at the workplace were reported. The exposure levels		
	of the workers were graded based on their jobs (Davies		
	1979; Davies 1984).		
	Historical cohort study among 666 men employed for at		
	least 1 year during 1950 to 1975 in five Japanese		
	chromate pigment production factories where lead		
	chromate, zinc chromate, molybdate orange and/or		
	strontium chromate were produced.		
	Sodium dichromate, anhydrous chromic acid and		
	potassium dichromate were used as the raw materials.		
	The follow-up study was ended in 1989. Three subjects		
	died from lung cancer, two of whom smoked. The SMR		
	results showed no excess mortality due to lung cancer or		
	other malignant neoplasmas among the workers engaged		
	in the manufacture of chromate pigment in Japan (O/E =		
	3/2.95). The actual exposure level at the work places was		
	unknown; however, the mean level of CrVI at the work		
	places after 1976 was 0.003–0.019 mg/m ³ (Kano et al. 1993).		
	1775).		
	Prospective cohort study among 251 men who worked at		
	a French lead and zinc chromate pigment factory for		
	more than 6 months from 1958 to 1977. The SMR for		
	lung cancer was 461 (95% CI = 270-790). The follow-up		
	study included 294 men who worked at this factory for at		
	least 6 months from 1958 to 1987. The SMR for lung		

Endpoint	Lowest effect levels/Results ²		
CASRN	1344-37-2 (C.I. Pigment Yellow 34; Chrome orange)	7758-97-6 (lead chromate)	7446-14-2 (lead sulfate)
	cancer was 360 (95% CI = 213-568); for digestive tract		
	cancer was 130 (95% CI = 60-247); for brain cancer		
	(only two cases) was 844 (95% CI = 102-3049).		
	Atmospheric samples taken in 1981 showed the		
	chromates level in the filtration department was in the		
	range of $2-3 \mu g/m^3$, in the grinding department, it was in		
	the range of $6-165 \mu g/m^3$, in the drying and sacking department, it was in the range of $6-178 \mu g/m^3$ and in the		
	sacks marking department, it was more than 2000 µg/m ³		
	(Haguenoer et al. 1981; Deschamps et al. 1995).		
	(11aguenoei et al. 1761, Deschamps et al. 1993).		
	Prospective cohort study among 1396 men who worked		
	for more than 6 months during 1945–1976, at three		
	German and two Dutch plants where zinc and lead		
	chromate pigments were produced. The SMR for lung		
	cancer was significantly increased in a restricted cohort		
	with 978 men employed before 1965 (SMR= 204; 95%		
	CI = 123-319). In the follow-up study, 1417 workers with		
	at least 1 year of exposure experience at the two German		
	plants were followed until 1988. The work place		
	exposure levels were reduced owing to the change of		
	production process. The SMR for bronchial carcinoma		
	was 227 (95% CI = 178-285) in the group of 739 workers		
	who worked before and after the change-over periods. In		
	the group of 678 workers who only worked after the change-over period, the SMR for lung cancer was 126		
	(95% CI = 58-238). The mean annual exposure level		
	during 1977 to 1987 was in the range of 0.012		
	-0.073 mg Cr/m ³ (Fentzel-Beyme 1983; Korallus et al.		
	1993).		
	Lead chromate pigment use		
	Historical cohort study among 226 white male spray-		
	painters from ten U.S. automobile assembly plants and		
	died during 1970 to 1976 or 1972 to 1976. The		

Endpoint	Lowest effect levels/Results ²		
CASRN	1344-37-2 (C.I. Pigment Yellow 34; Chrome orange)	7758-97-6 (lead chromate)	7446-14-2 (lead sulfate)
	proportionate mortality ratio (PMR) for respiratory cancer among the spray-painters was increased, but did not reach statistical significance when compared with the country and non-spray painter reference groups (PMR = 139 and 108, respectively). In a nested case-control study, the relative risk for lung cancer associated with spray-painting experience was analysed. 263 decedents who died from lung cancer were identified in the ten plants. 1001 controls were decedents who died from circulatory disease or accidents from the same population (matched for plant and age). Mantel-Haenszel odd ratio analysis showed the relative risk for lung cancer in the white male spray-painters was not significantly increased. The paints contained chromate pigments, no further exposure information or employment history data available (Chiazze et al. 1980).		
	Additional studies: Dalager et al. 1980, Langård and Norseth 1975, and Langård and Vigander 1983: workers were mainly exposed to zinc chromate pigments. Bertazzi et al. 1981: workers were co-exposed to asbestos, zinc, barium and strontium chromates. Chen and Seaton 1998: literature-based meta analysis among painters; workers were exposed to organic solvents, lead chromate pigment, asbestos and others.		
Reproductive and developmental toxicity	No data identified.	Retrospective survey among workers in a lead chromate production plant in China. 22 exposed female workers with 42 controls and 19 exposed male worker's wives with 91 controls were enrolled in this study. The rates for threatened abortion were significantly increased among the female workers and the wives of male workers exposed to lead chromate (31.8% in exposed women versus 2.4 % in controls and 36.8% in	No data identified.

Endpoint	Lowest effect levels/Results ²		
CASRN	1344-37-2 (C.I. Pigment Yellow 34; Chrome orange)	7758-97-6 (lead chromate)	7446-14-2 (lead sulfate)
		exposed men's wives versus 2.2% in controls,	
		respectively), no further information available	
		(Wang and Zhao 1985).	

- 1. The toxicity data for lead chromate (7758-97-6) and lead sulfate (7446-12-2) are included in the assessment for C.I. Pigment Yellow 34 (1344-37-2) as they are the principal components of the latter.
- 2. LC₅₀ = median lethal concentration; LD₅₀ = median lethal dose; LD_{LO} = lowest lethal dose; LOEC = Lowest-Observed-Effect Concentration; LOEL = Lowest-Observed-Effect Level.