Screening Assessment for the Challenge

Acetamide, N-[5-[[2-(acetyloxy)ethyl](phenylmethyl)amino]-2-[(2,4-dinitrophenyl)azo]-4-methoxyphenyl]-

Chemical Abstracts Service Registry Number 16421-41-3

Environment Canada Health Canada

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Synopsis

Pursuant to section 74 of the *Canadian Environmental Protection Act, 1999* (CEPA 1999), the Ministers of the Environment and of Health have conducted a screening assessment on Acetamide, *N*-[5-[[2-(acetyloxy)ethyl](phenylmethyl)amino]-2-[(2,4-dinitrophenyl)azo]-4-methoxyphenyl]- (AADM), Chemical Abstracts Service Registry Number 16421-41-3. This substance was identified as a high priority for screening assessment and included in the Challenge because it had been found to meet the ecological categorization criteria for persistence, bioaccumulation potential and inherent toxicity to non-human organisms and is believed to be in commerce in Canada.

The substance AADM was not considered to be a high priority for assessment of potential risks to human health, based upon application of the simple exposure and hazard tools developed by Health Canada for categorization of substances on the *Domestic Substances List*. Therefore, this assessment focuses on information relevant to the evaluation of ecological risks.

AADM is an organic substance that was previously used in Canada as a colourant dye in textiles and fabric based on use codes from the 1986 *Domestic Substances List*. The substance is not naturally produced in the environment. No companies reported manufacturing, importing or using this substance in Canada above the reporting thresholds in 2005 or 2006. However the threshold of 100 kg was used throughout this screening assessment to capture the maximum potential mass of this substance in use in Canada.

Based on reported use patterns for similar disperse azo dyes and on certain assumptions, most of the substance, when used, is expected to end up in solid waste disposal sites but a significant proportion is estimated to be released to sewer water (14.8%). AADM is not expected to be soluble in water or to be volatile; it is expected to partition to particles because of its hydrophobic nature. For these reasons, after release to water, AADM will likely end up mostly in sediments, and to a lesser extent, in agricultural soil that has been amended with sewage sludge. It is not expected to be significantly present in other media. It is also not expected to be subject to long-range atmospheric transport.

Based on its physical and chemical properties, AADM is expected to be persistent in the environment (in water, sediment and soil). However, new experimental data relating to the bioaccumulation potential of a relatively close structural analogue of AADM suggest that this dye has a low potential to accumulate in the lipid tissues of organisms. The substance meets the persistence criteria but does not meet the bioaccumulation criteria as set out in the *Persistence and Bioaccumulation Regulations*. In addition, experimental toxicity data for a chemical analogue suggest that the substance does not harm aquatic organisms exposed to low concentrations.

For this screening assessment, two conservative exposure scenarios were selected in which an industrial operation (user of the dye) and consumer use of products containing this substance resulted in discharges of AADM into the aquatic environment. The predicted environmental concentrations in water were below the predicted no-effect concentration calculated for sensitive aquatic organisms.

This substance will be included in the upcoming Domestic *Substances List* inventory update initiative. In addition and where relevant, research and monitoring will support verification of assumptions used during the screening assessment.

Based on the information available, it is concluded that AADM does not meet any of the criteria set out in section 64 of CEPA 1999.

Introduction

The Canadian Environmental Protection Act, 1999 (CEPA 1999) (Canada 1999) requires the Minister of the Environment and the Minister of Health to conduct screening assessments of substances that have met the categorization criteria set out in the Act to determine whether these substances present or may present a risk to the environment or human health. Based on the results of a screening assessment, the Ministers can propose to take no further action with respect to the substance, to add the substance to the Priority Substances List (PSL) for further assessment, or to recommend that the substance be added to the List of Toxic Substances in Schedule 1 of the Act and, where applicable, the implementation of virtual elimination.

Based on the information obtained through the categorization process, the Ministers identified a number of substances as high priorities for action. These include substances that

- met all of the ecological categorization criteria, including persistence (P), bioaccumulation potential (B) and inherent toxicity to aquatic organisms (iT), and were believed to be in commerce in Canada; and/or
- met the categorization criteria for greatest potential for exposure (GPE) or
 presented an intermediate potential for exposure (IPE), and had been identified as
 posing a high hazard to human health based on classifications by other national or
 international agencies for carcinogenicity, genotoxicity, developmental toxicity or
 reproductive toxicity.

The Ministers therefore published a notice of intent in the *Canada Gazette*, Part I, on December 9, 2006 (Canada 2006a), that challenged industry and other interested stakeholders to submit, within specified timelines, specific information that may be used to inform risk assessment, and to develop and benchmark best practices for the risk management and product stewardship of these substances identified as high priorities.

The substance AADM was identified as a high priority for assessment of ecological risk as it had been found to be persistent, bioaccumulative and inherently toxic to aquatic organisms and is believed to be in commerce in Canada. The Challenge for this substance was published in the *Canada Gazette* on February 16, 2008 (Canada 2008). A substance profile was released at the same time. The substance profile presented the technical information available prior to December 2005 that formed the basis for categorization of this substance. As a result of the Challenge, submissions of information pertaining to the bioaccumulation and toxicity (as analogues to this substance) were received.

Although AADM was determined to be a high priority for assessment with respect to the environment, it did not meet the criteria for GPE or IPE, and was not identified as posing a high hazard to human health based on classifications by other national or international

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agencies for carcinogenicity, genotoxicity, developmental toxicity or reproductive toxicity. Therefore, this assessment focuses principally on information relevant to the evaluation of ecological risks.

Screening assessments under CEPA 1999 focus on information critical to determining whether a substance meets the criteria for defining a chemical as toxic as set out in section 64 of the Act, where

- "64. [...] a substance is toxic if it is entering or may enter the environment in a quantity or concentration or under conditions that
 - (a) have or may have an immediate or long-term harmful effect on the environment or its biological diversity;
 - (b) constitute or may constitute a danger to the environment on which life depends; or
 - (c) constitute or may constitute a danger in Canada to human life or health."

Screening assessments examine scientific information and develop conclusions by applying a weight of evidence approach and precaution.

This screening assessment considers any new information on chemical properties, hazards, uses and exposure submitted under the Challenge. Data relevant to the screening assessment of this substance were identified in original literature, review documents, stakeholder research reports and from recent literature searches up to October 2008. Key studies were critically evaluated and generally only results from studies of good quality were used to reach conclusions, although other studies and modelling results may have been considered as part of the weight of evidence. When available and relevant, information presented in hazard assessments from other jurisdictions was also used. The screening assessment does not represent an exhaustive or critical review of all available data. Rather, it presents the most critical studies and lines of evidence pertinent to the conclusion.

This screening assessment was prepared by staff in the Existing Substances Program at Health Canada and Environment Canada and it incorporates input from other programs within these departments. The assessment has undergone external written peer review. While external comments were taken into consideration, the final content and outcome of the screening risk assessment remain the responsibility of Health Canada and Environment Canada. Additionally, the draft of this screening assessment was subject to a 60-day public comment period. The critical information and considerations upon which the assessment is based are summarized below.

Substance Identity

For the purposes of this document, the substance Acetamide, *N*-[5-[[2-(acetyloxy)ethyl](phenylmethyl)amino]-2-[(2,4-dinitrophenyl)azo]-4-methoxyphenyl] will be referred to as AADM which has been derived from the DSL name. Information on substance identity is included in Table 1.

Table 1. Substance Identity

| Chemical Abstracts Service Registry Number (CAS RN) | 16421-41-3 | | | | |
|--|---|--|--|--|--|
| DSL name | Acetamide, N-[5-[[2-(acetyloxy)ethyl](phenylmethyl)amino]-2-[(2,4- | | | | |
| | dinitrophenyl)azo]-4-methoxyphenyl]- | | | | |
| | Acetamide, N-[5-[[2-(acetyloxy)ethyl](phenylmethyl)amino]-2-[(2,4- | | | | |
| | dinitrophenyl)azo]-4-methoxyphenyl]- (TSCA, DSL, AICS, ASIA-PAC) | | | | |
| | 2-[[5-acetamido-4-[(2,4-dinitrophenyl)azo]-2- | | | | |
| Inventory names ¹ | methoxyphenyl]benzylamino]ethyl acetate (EINECS) | | | | |
| | 2-[[5-Acetamido-4-[(2,4-dinitrophenyl)azo]-2- | | | | |
| | Acetamide, N-[5-[[2-(acetyloxy)ethyl](phenylmethyl)amino]-2-[(2,4- | | | | |
| | dinitrophenyl)azo]-4-methoxyphenyl]- (PICCS) | | | | |
| 041 | p-Acetanisidide, 5'-[benzyl(2-hydroxyethyl)amino]-2'-[(2,4- | | | | |
| Other names | dinitrophenyl)azo]-, acetate (ester) | | | | |
| Chemical group | Discrete organics | | | | |
| Chemical sub-group | Monoazo dye | | | | |
| Chemical formula | $C_{26}H_{26}N_6O_8$ | | | | |
| Chemical structure | | | | | |
| SMILES ² | O=C(OCCN(c(c(OC)cc(N=Nc(c(N(=O)(=O))cc(N(=O)(=O))c1)c1)c2NC(=O) C)c2)Cc(cccc3)c3)C | | | | |
| Molecular mass | 550.53 g/mol | | | | |
| | <u> </u> | | | | |

National Chemical Inventories (NCI). 2006: AICS (Australian Inventory of Chemical Substances); ASIA-PAC (Combined Inventories of Asia-Pacific Region); EINECS (European Inventory of Existing Chemical Substances); PICCS (Philippine Inventory of Chemicals and Chemical Substances); and TSCA (Toxic Substances Control Act Chemical).

² Simplified Molecular Line Input Entry System

Physical and Chemical Properties

No experimental data are available for AADM. At the Environment Canada-sponsored Quantitative Structure-Activity Relationship (QSAR) Workshop in 1999 (Environment Canada 2000), modeling experts identified many structural classes of pigments and dyes as "difficult to model" using QSARs. The physical and chemical properties of many of the structural classes of dyes and pigments (including acid and disperse dyes) are not amenable to model prediction because they are considered "out of the model domain of applicability" (e.g., structural and/or property parameter domains). Therefore, to determine potential utility, the domains of applicability of QSAR models to dyes and pigments are evaluated on a case-by-case basis. It is generally considered inappropriate to use QSAR models to predict the physical and chemical properties of AADM. Consequently, a number of analogues were identified and "read-across" data has been used to determine the approximate physical and chemical properties in Table 2. These properties were subsequently used for further modeling and lines of evidence in this assessment.

An analogue is a chemical which is structurally similar to the substance under assessment and is therefore expected to have similar physical-chemical properties, behaviour in the environment and/or toxicity. Where there are experimental data for a given parameter for an analogue substance, these can be used directly or with adjustment as an estimate of that parameter value for the substance under assessment.

In order to find acceptable analogues, a review of data for several disperse azo dyes was performed (Anliker *et al.* 1981, Anliker and Moser 1987, Baughman and Perenich 1988, ETAD 1995, Brown 1992, Yen *et al.* 1989, Sijm *et al.* 1999). These compounds have structural similarities to AADM but also share other important attributes that contribute to their suitability as analogues. This includes properties affecting their fate in the environment such as high molecular weights (generally >300 g/mol), similar cross sectional diameters (1.43 – 2.05 nm), solid particulate structures, decomposition at greater than 74 °C (to 240 °C), and "dispersibility" in water (i.e. not truly soluble). In addition, they have limited solubility in n-octanol, a negligible vapour pressure and are stable under environmental conditions as they are designed to be so.

Table 2 contains analogue as well as read-across experimental and modelled physical-chemical properties of AADM that are relevant to its environmental fate. No experimental values were found for AADM.

Table 2. Physical and chemical properties for AADM and relevant chemical analogues.

| Property | Type ¹ | Value | Temperature (°C) | Reference | | | |
|---|---|--|------------------|---|--|--|--|
| | Analogue Disperse Blue 79 | 157 | | PhysProp 2006 | | | |
| Melting point ² (°C) | Read-across for disperse azo dyes | 117 to 175 74 to 236 | | Anliker and Moser 1987, Baughman and Perenich 1988 | | | |
| | Analogue Disperse Blue 79:1 | ≥138-153 | | Sandoz Chemicals 1989, Yen <i>et al</i> . 1989 | | | |
| Boiling point ³ (°C) | | Not Applicable | | | | | |
| Density (kg/m³) | | Not A | vailable | | | | |
| Vapour pressure | Analogue Disperse Blue 79 | 4.53x10 ⁻⁷ | | Clariant 1996 | | | |
| (Pa) | Read-across for disperse azo dyes | $5.33 \times (10^{-12} \text{ to} $ $10^{-5})$ $(4x10^{-14} \text{ to } 4 \times $ $10^{-7} \text{ mm Hg})$ | 25 | Baughman and Perenich 1988 | | | |
| Henry's Law constant (Pa·m³/mol) | Read-across ⁴ | 10 ⁻⁸ to 10 ⁻¹ (10 ⁻¹³ to 10 ⁻⁶ atm·m ³ /mol) | | Baughman and Perenich 1988 | | | |
| Log K _{ow} (Octanol-water partition coefficient) (dimensionless) | Analogue Disperse Blue 79 | 4.1, 4.4 | | Clariant 1996, Brown 1992 | | | |
| (difficilisioniess) | Analogue Disperse Blue | 4.44, 4.8 | | Sijm <i>et al</i> . 1999, Yen <i>et al</i> . | | | |

| Property | Type ¹ | Value | Temperature (°C) | Reference |
|---|---|---|------------------|--|
| | 79:1 | | | 1989 |
| | Read-across for disperse azo dyes | 1.79 to 5.1 | | Baughman and Perenich 1988 |
| | Read-across for disperse azo dyes | >2 -5.1 | | Anliker <i>et al.</i> 1981; Anliker and Moser 1987 |
| | Analogue Disperse Orange 30 | 4.2 | | Brown 1992 |
| Log K _{oc} (Organic carbon partition coefficient) (dimensionless) | Read-across, calculated ⁵ | 3.4 to 4.2 | | Baughman and Perenich 1988 |
| Water solubility (mg/L) | Analogue Disperse Blue 79 | 0.0054, 0.02, 0.000938 | 15-25 | Clariant 1996, Brown 1992, Baughman and Perenich 1988 |
| | | <0.01 | | Anliker and Moser 1987 |
| | Read-across for disperse azo dyes | 1.2×10^{-5} to $35.5 (4 \times 10^{-11}$ to 1.8×10^{-4} mol/L) | | Baughman and Perenich 1988 |
| | | substantially water insoluble | | ETAD 1995 |
| | Analogue Disperse Blue 79:1 | 0.0052 | 25 | Baughman and Perenich 1989 |
| | Analogue Disperse | 0.07 | | Brown 1992 |

| Property | Type ¹ | Value | Temperature (°C) | Reference |
|--|---|--|---------------------|--------------------------------|
| | Orange 30 | | | |
| n-octanol solubility (mg/L) | Read-across for disperse azo dyes | 81-2100 | 20 | Anliker and Moser 1987 |
| pK _a (Acid dissociation constant) (dimensionless) | Modelled | 13.3 for acid form 0.87 for base form | | ACD/pK _a DB 2005 |

¹ These extrapolated values used for AADM are based on evidence on disperse dyes submitted to Environment Canada under the New Substance Notification Regulations (ETAD 1995) and available evidence from other disperse dye analogues found in literature.

² The phrase melting point is used but this may be better referred to as a decomposition point because disperse dyes are known to char at high temperatures (greater than 200°C) rather than melt.

Structural disperse azo analogues to AADM are presented in Table 3a and 3b below. Certain physical and chemical properties (see Table 2), empirical bioaccumulation data (Table 6) and empirical toxicity data (see Table 7) of these analogues were used in support of the weight of evidence and decisions in this SAR. Specifically, data were obtained for the structural analogues: Disperse Orange 30, Disperse Blue 79, Disperse Blue 79:1, Disperse Red 73, Disperse Orange 25, Disperse Red 17 and Disperse Yellow 3 (Table 3a).

Table 3a. Structural analogues for AADM.

| | CAS RN | Common Name | DSL name ¹ | Structure of analogue | Available empirical data |
|----|-----------|--------------------|--|-----------------------|---|
| i. | 5261-31-4 | Disperse Orange 30 | Propanenitrile, 3-[[2-(acetyloxy)ethyl][4-[(2,6-dichloro-4-nitrophenyl)azo]phenyl] amino]- | N C CH ₃ | Bioaccumul ation, log k _{ow} , water solubility |

³ Boiling point is generally not applicable for disperse dyes. For powder dyes, charring or decomposition occurs at high temperatures instead of boiling. For liquids and pastes, boiling will only occur for the solvent component while the unevaporated solid will decompose or char (ETAD 1995).

⁴ Solubilities of several disperse dyes at 25 and 80°C were used by Baughman and Perenich (1988) to calculate Henry's Law constants for these dyes. These values are presented here as a range to illustrate the expected Henry's Law constant for AADM.

⁵ Log K_{oc} values are based on calculations by Baughman and Perenich (1988) using a range of measured solubilities for commercial dyes and an assumed melting point of 200°C.

| ii | 12239-34-8 | Disperse Blue 79 | Acetamide, N-[5-[bis[2-(acetyloxy)ethyl]amino] -2-[(2-bromo-4,6-dinitrophenyl)azo]-4-ethoxyphenyl]- | H ₃ C CH ₃ H ₃ C CH ₃ H ₃ C CH ₃ | Melting point, vapour pressure, log kow, water solubility, aquatic toxicity, |
|------|------------|-----------------------|--|--|--|
| iii. | 3618-72-2 | Disperse Blue 79:1 | Acetamide, N-[5-[bis[2-(acetyloxy)ethyl] amino]-2-[(2-bromo-4,6-dinitrophenyl)azo]-4-methoxyphenyl]- | H ₃ C OH ₃ H ₃ C OH ₃ | Melting point, log K _{ow} , water solubility, aquatic toxicity |
| iv. | 16889-10-4 | Disperse Red 73 | 2-((4-((2-Cyanoethyl) ethylamino)phenyl)azo) -5-nitro benzonitrile | | Aquatic toxicity |
| V. | 31482-56-1 | Disperse Orange 25 | 3-(Ethyl(4-((4-nitro phenyl)azo)phenyl) amino)propanenitrile | N = N | Aquatic toxicity |
| vi. | 3179-89-3 | Disperse Red 17 | Ethanol, 2,2'-((3-methyl-4-(2-(4-nitrophenyl) diazenyl)phenyl) imino)bis- | H ₁ C HO | Aquatic toxicity |
| vii | 2832-40-8 | Disperse Yellow 3 | 4-(2-Hydroxy-5- methylphenylazo)aceta nilide | OH3 OH CH3 | Aquatic toxicity |

It should be noted that there are several uncertainties associated with the use of physical and chemical-toxicological and bioaccumulation data available for the substances presented in Table 3a. All these substances belong to the same chemical class (disperse azo dyes with their characteristic azo bond) and are used for similar industrial purposes However, there are differences between these substances associated with their unique functional groups (see Table 3b below). As a result, these analogues have empirical water solubilities that range over four orders of magnitude from 10⁻⁵ to 0.07 mg/L (see Table 2 above). Due to this variability, caution should be exercised when drawing conclusions from these values. However the analogue data is presented to be considered into the weight of evidence for this substance.

Table 3b. Comparisons of structural analogues with AADM.

| | CAS RN | Common Name | Molecular | % | Minimum-maximum cross-sectional |
|-----|------------|--------------------|----------------|-----------------------------------|---------------------------------|
| | | | mass (g/mol | structure similar ¹ | diameter ² (nm) |
| i | 3618-72-2 | Disperse Blue 79:1 | 625.39 | 91.43 | 1.43-2.03 |
| ii | 12239-34-8 | Disperse Blue 79 | 639.42 | 88.28 | 1.69-2.045 |
| iii | 5261-31-4 | Disperse Orange 30 | 450.28 | 68.29 | 1.75-1.98 |
| iv. | 16889-10-4 | Disperse Red 73 | 348.36 | NA | 1.31-1.93 |
| V. | 31482-56-1 | Disperse Orange 25 | 323.35 | NA | 1.37-1.95 |
| vi. | 3179-89-3 | Disperse Red 17 | 344.36 | NA | 1.41-1.86 |
| vii | 2832-40-8 | Disperse Yellow3 | 269.31 | NA | 1.59-1.70 |

¹ ChemID Plus 2008, value presented if >60%

² CPOP (2008)

Sources

AADM is not naturally produced in the environment.

Recent information was collected through industry surveys conducted for the years 2005 and 2006 under Canada Gazette Notices issued pursuant to section 71 of CEPA 1999 (Canada 2006b and 2008). These Notices required submission of data on the Canadian manufacture and import of the substance. For 2006, data were also required on use quantities of AADM.

In 2006, no companies reported importing or manufacturing AADM above the prescribed reporting threshold of 100 kg/year in Canada. No companies reported using a total quantity greater than 1000 kg of the substance, whether alone, in a mixture, in a product or in a manufactured item, at any concentration in 2006. In the Declaration of Stakeholder-Interest form associated with the section 71 survey for 2006, one company reported a stakeholder interest in this substance despite not meeting mandatory reporting requirements (Canada 2008).

In 2005, no companies reported manufacturing or importing AADM in quantities above the prescribed reporting threshold of 100 kg/year. However, one company identified themselves as having a stakeholder interest in the substance (Canada 2006b).

The quantity reported during development of the *Domestic Substances List* (DSL) to be manufactured, imported or in commerce in Canada during the calendar year 1986 was 100 kg (Environment Canada 1988). The number of notifiers for the calendar years 1984-86 was fewer than four.

AADM is an existing chemical in Europe, but is not on the low or high production volume chemicals lists (ESIS 2008). The Substances in Preparation in Nordic Countries (SPIN) database states that this substance was in use in Denmark and Sweden from 1999 to 2006, but the quantities in use in those countries were not stated (SPIN 2008).

No manufacturing, importation or use of this substance was reported for 2006. However the s. 71 reporting threshold (i.e., 100 kg) was used throughout this screening assessment to capture the maximum potential mass of this substance in use in Canada that would not otherwise be subject to reporting.

Uses

No recent information on the use of this substance in Canada has been identified. The following DSL use codes were identified for the substance during the DSL nomination (1984-1986): "Colourant - pigment/stain/dye/ink" and "Textile, Primary Manufacture" (Environment Canada 1988). No additional information on potential uses of AADM was identified through searches of the available scientific and technical literature.

Releases to the Environment

Mass Flow Tool

To estimate potential releases of the substance to the environment at different stages of its life cycle, the Mass Flow Tool was developed (Environment Canada 2008a). Empirical data concerning releases of specific substances to the environment are seldom available. Therefore, for each identified type of use of the substance, the proportion and quantity of release to the different environmental media are estimated, as are the proportion of the substance chemically transformed or sent for waste disposal. Unless specific information on the rate or potential for release of the substance from landfills and incinerators is available, the Mass Flow Tool does not quantitatively account for releases to the environment from disposal.

Assumptions and input parameters used in making the release estimates are based on information obtained from a variety of sources including responses to regulatory surveys, Statistics Canada, manufacturers' websites and technical databases and documents. Of particular relevance are emission factors, which are generally expressed as the fraction of a substance released to the environment, particularly during its manufacture, processing, and use associated with industrial processes. Sources of such information include emission scenario documents, often developed under the auspices of the Organization for Economic Cooperation and Development (OECD), and default assumptions used by different international chemical regulatory agencies. It is noted that the level of uncertainty in the mass of substance and quantity released to the environment generally increases towards the end of the life-cycle.

The Mass Flow Tool result for other in-commerce disperse azo dyes was used in this document to estimate the fraction of AADM that could be released to the environment, since AADM is structurally similar to other disperse azo dyes and their use patterns are also expected to be similar (textiles).

Based on Statistics Canada information and an analysis by Industry Canada (2008), it is proposed that AADM may be imported in manufactured articles. A ratio of textiles manufactured in Canada / imported textiles of 30/70 has been used to estimate the amount of dye imported in textiles (Environment Canada 2008b). This import quantity was included in the Mass Flow Tool calculations as well as in the exposure scenarios developed.

Table 4. Estimated releases and losses of disperse azo dyes to environmental media, chemical transformation and transfers to waste disposal sites, based on the Mass Flow Tool.

| Fate | Proportion of the | Major life cycle stage |
|------|-------------------|------------------------|
|------|-------------------|------------------------|

| | $\operatorname{mass}\left(\%\right)^{1}$ | involved ² |
|---------------------------------|--|-----------------------------|
| Releases to receiving media: | | |
| To soil | 0.0 | n/a^3 |
| To air | 0.0 | n/a |
| To sewer ⁴ | 14.8 | Formulation, consumer use |
| Chemically transformed | 0.0 | n/a |
| Transferred to waste | 85.2 | Formulation, waste disposal |
| disposal sites (e.g., landfill, | | |
| incineration) | | |

¹ For disperse azo dyes, information from the following OECD emission scenario documents was used to estimate releases to the environment and distribution of the substance as summarized in this table: OECD 2004, 2007. Values presented for release to environmental media do not account for possible mitigation measures that may be in place in some locations (e.g., partial removal by sewage treatment plants). Specific assumptions used in derivation of these estimates are summarized in Environment Canada 2008b

Results indicate that AADM can be expected to be found largely in waste management sites (85.2%), due to the eventual disposal of manufactured items containing it. Mass Flow Tool calculations do not quantitatively account for releases of the substance to the environment from waste disposal sites (such as landfills, incinerators) unless specific information on the rate or potential for release is available. No such information has been identified for AADM. A small fraction of solid waste is incinerated which is expected to result in transformation of the substance. Based largely on information contained in OECD emission scenario documents for processing and uses associated with this type of substance, it is estimated that 14.8% of AADM may be released to sewers.

Based on the above, sewer water is the medium receiving the greatest proportion of AADM emitted during product processing. It is anticipated that the majority of the substance bound in products will be sent to landfills for disposal.

Environmental Fate

As indicated by the results of the Mass Flow Tool (Table 4), the substance AADM is expected to be released to waste water effluents during industrial processing and use. The moderate to high log K_{ow} (analogues 4.1 to 4.8) and high log K_{oc} (read-across: 3.4 to 4.2) values (see Table 2) indicate that this substance may have affinity for solids. However, the log K_{oc} is a calculated value (see footnote 3 below Table 2) and the adsorption potential of solid particulate dye structures is generally not well understood, therefore the degree of adsorption of AADM is uncertain.

AADM is expected to be mostly found in sediment or soil, and is not expected to be subject to long-range atmospheric transport.

² Applicable stage(s): production-formulation-industrial use-consumer use-service life of article/product-waste disposal.

³ Not applicable

⁴ Wastewater before any form of treatment

AADM does not biodegrade fast (see Table 5 below). It may inadvertently be applied to agricultural soils and pasture lands in Canada as a component of biosludge which is commonly used for soil enrichment. Moreover, it may also be released from coloured textiles deposited in landfills.

In solution AADM can behave as either an acid or as a base. With an estimated pK_a for the acid of 13.3 and for the base of 0.87 (see Table 2) dissolved forms of AADM are not expected to ionize in water at environmentally relevant pHs. Since disperse azo dye analogues have shown limited water solubility (see Table 2) AADM is expected to be only sparingly soluble. Because of its low solubility, when released into water, this substance is behave as a colloidal dispersion (Yen et al. 1991). Thus, this substance will expected to be mostly present as a solid or adsorbed to suspended particles and sink eventually to bed sediments where it is expected to remain in a relatively biologically unavailable form. It has been concluded by Yen at al. (1989) that disperse dyes tend to accumulate extensively in sediments and biota unless they are degraded at rates comparable to uptake. Razo-Flores et al. (1997) have stated that due to the recalcitrant nature of azo dyes in the aerobic environment, they eventually end up in anaerobic sediments, shallow aguifers and in groundwater. Yen et al. (1991) observed that some azobenzene dye analogues were transformed under anaerobic conditions in sediment via hydrolysis and reduction, and concluded that most azo dyes would likely not persist in anaerobic sediment systems. In buried sediment AADM may undergo anaerobic degradation, as described in the following section on Persistence.

The rate of volatilization from water is proportional to the Henry's law constant (Baughman and Perenich 1988). The low to negligible Henry's Law constant (10^{-8} to 10^{-1} atm·m³/mol, read-across data in Table 2) and the low to negligible vapour pressure (5.33 × (10^{-12} to 10^{-5}) Pa, read-across data in Table 2) indicate that AADM is essentially non-volatile. Therefore, volatilization is not likely to be an important transport pathway for the loss of this substance from moist and dry soil surfaces nor from aquatic compartments. Baughman and Perenich (1988) also state that volatilization will not be an important transport pathway for the loss of disperse dye from aquatic systems. This is consistent with the physical state (solid particulate structure) of AADM which does not make it a likely candidate for volatilization.

Persistence and Bioaccumulation Potential

Persistence

No experimental biological degradation data for AADM have been identified. According to the Ecological and Toxicological Association of Dyes and Organic Pigments Manufacturers, with some exceptions, dyes are considered essentially non-biodegradable under aerobic conditions (ETAD 1995). Repeated evaluation of ready and inherent biodegradability using accepted screening tests (see OECD Guidelines for Testing Chemicals) have generally confirmed this expectation (Pagga and Brown 1986; ETAD

1992). Based on the chemical structure of AADM, there is no reason to suspect that biodegradation will be other than that described for dyes generally (ETAD 1995).

Some disperse azo dyes have been shown to undergo relatively rapid anaerobic degradation in sediment at depth where anoxic conditions persist (Yen *et al.* 1991, Baughman and Weber 1994, Weber and Adams 1995). Disperse dyes enter the aquatic system mostly as a dispersion of fine suspended particles, eventually settling to the aerobic layers of surface sediment where they will persist until sediment burial creates reducing conditions. The rate of sediment deposition and the extent of bioturbation varies from site to site and thus it is very difficult to ascertain the residence time of dyes in aerobic sediment layers. It is likely however, that in many cases this is greater than 365 days. Once under anaerobic or reducing conditions, azo dyes may undergo rapid degradation to substituted aromatic amine constituents as demonstrated by Yen *et al.* (1991) who measured reduction half-life values in compacted sediments at room temperature of 2.9 to 2.0 days for azobenzene dyes. However, in deep anoxic sediment, these biodegradation transformation products are not expected to present a high degree of exposure potential to most aquatic organisms, and therefore they are not likely to present an ecological concern.

Since no experimental biodegradation data are available for AADM, a QSAR-based weight-of-evidence approach (Environment Canada 2007) was applied using the degradation models shown in Table 5 below. Although the expected release of AADM will be to wastewater, its residence time in the water column may be short before finally sinking to the sediment bed due to its low solubility and behaviour as a colloidal dispersion. However, given the lack of data regarding this issue, persistence was primarily examined using predictive QSAR models for biodegradation in water. The following analysis applies primarily to the portion of this substance that is present in the environment in the dissolved form, recognizing that a significant proportion would also likely exist in dispersed form as solid particles. AADM does not contain functional groups expected to undergo hydrolysis in aerobic environments (dyes are designed to be stable in aqueous conditions). Table 5 summarizes the results of available QSAR models for biodegradation in water.

Table 5. Modelled data for biodegradation of AADM

| Model | Model Basis | Medium | Value | Interpretation | Extrapolated half-life (days) | Extrapolation Reference and/or Source |
|-------------------------|---|--------------------|--------|--------------------------------|-------------------------------|---|
| BIOWIN1* v4.1 (2000) | Linear probability | water (aerobic) | 0.0726 | Does not biodegrade fast | n/a | |
| BIOWIN2* v4.1 (2000) | Non-linear probability | water (aerobic) | 0.00 | Does not biodegrade fast | n/a | |
| BIOWIN3* v4.1 (2000) | Expert Survey (ultimate biodegradation) | water (aerobic) | 1.13 | Recalcitrant | 180 | US EPA 2002 |
| BIOWIN4* v4.1 (2000) | Expert Survey (primary | water (aerobic) | 3.01 | Weeks | 15 | US EPA 2002 |

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| | biodegradation) | | | | | |
|--|-----------------------------------|--------------------|-------|--------------------------------|-------|---------------------|
| BIOWIN5* v4.1 (2000) | MITI linear probability | water (aerobic) | -0.69 | Does not biodegrade fast | n/a | |
| BIOWIN6* v4.1 (2000) | MITI non-linear probability | water (aerobic) | 0.00 | Does not biodegrade fast | n/a | |
| BIOWIN Overall Conclusion ¹ | BIOWIN 3 + BIOWIN 5 | water (aerobic) | no | Not readily biodegradable | n/a | |
| CATABOL v. 5.10.2 | % BOD (OECD 301C) | water (aerobic) | 19 | Persistent (<20%) | > 182 | Aronson et al. 2006 |

^{*}BIOWIN 1–6 are outputs obtained from the predictive model BIOWIN (2000). BIOWIN estimates aerobic biodegradability of organic chemicals using six different models.

The results from Table 5 show that the majority of the probability models (BIOWIN 1, 2, 5, 6) suggest this substance does not biodegrade rapidly. In fact, all probability results are less than 0.3, the cut-off suggested by Aronson *et al.* (2006) identifying substances as having a half-life >60 days (based on the MITI probability models). The half-life from the primary survey model (BIOWIN 4) result of weeks is suggested to mean approximately 15 days (US EPA 2002, Aronson et al 2006); however, the nature of the degradation products is unknown. The ultimate survey model (BIOWIN 3) result of recalcitrant is suggested to mean 180 days by the US EPA (2002). The overall conclusion from BIOWIN (2000) is that this substance is not readily biodegradable.

CATABOL (c2004–2008) predicted 19 % rate of biodegradation based on the OECD 301 readily biodegradation test (%BOD) which has been suggested as meaning likely persistent (Aronson and Howard 1999) and having a half-life in water of >182 days.

When the results of the probability models, the overall BIOWIN conclusion and ultimate degradation models are considered, there is model consensus suggesting that the half-life in water is >182 days, which is consistent with what would be expected for a chemical used as a disperse dye (i.e., manufactured to be relatively insoluble and durable). Using a ratio of 1:1:4 for a water:soil:sediment half-life extrapolation ratio (Boethling 1995), the half-life in soil should be >182 days and the half-life in aerobic sediments should be >365 days.

Based on the results of predictive modelling (principally for ultimate degradation) and on expert judgement (ETAD 1995), AADM meets the persistence criteria for water and soil (half life in soil and water ≥ 182 days) as well as sediments (half life in sediments ≥ 365 days) as set out in the *Persistence and Bioaccumulation Regulations* (Canada 2000).

Potential for Bioaccumulation

¹Based on outcome of BIOWIN 3 and BIOWIN 5.

No experimental bioaccumulation data are available for AADM.

In the absence of experimental and modelled data for AADM itself, bioconcentration (BCF) and bioaccumulation (BAF) factors for structural analogues were used to estimate AADM's potential for bioaccumulation. To that end, a bioconcentration study submitted for a relatively close structural analogue, Disperse Orange 30, suggests that it is unlikely to accumulate in fish (Shen and Hu 2008). This test was performed according to OECD Guidelines for Testing of Chemicals, Test No. 305B-1996, Bioconcentration: Semi-Static Fish Test. The bioconcentration of Disperse Orange 30 in zebra fish (*Brachydanio rerio*) was determined in a 28-day semi-static test with a test medium renewal every two days. An exposure test at a nominal concentration of 20 mg/L (mean measured concentration 0.028 - 0.28 mg/L) was performed in accordance with the result of the fish acute toxicity test to check the bioconcentration potential of the test substance. Samples from both test solutions and test organisms were taken daily from the 26th day to the last day during the 28-day exposure test period. Samples were prepared by extracting the lipid component from the test fish. The measured concentration of test substance, fish lipid content and BCF calculation are reported in Table 6.

Table 6. Measured concentration of Disperse Orange 30, fish lipid content and BCF calculation

| Treatments (20 mg/L) | 5 | Sampling Time | |
|--|--------------------------|--------------------------|--------------------------|
| | The 26 th day | The 27 th day | The 28 th day |
| Measured concentration of the test substance in extracted solutions (mg/L) | <0.028 | <0.028 | <0.028 |
| Content of the test substance in the fish lipids (mg) | <1.68 | <1.68 | <1.68 |
| Fish total weight (g) | 2.07 | 2.13 | 2.53 |
| Concentration of the test substance in the fish C _f (mg/kg) | < 0.81 | < 0.79 | < 0.66 |
| Measured concentration of the test substance in the water $C_{\rm w}$ (mg/L) | 0.028 ~ 0.28 | 0.028 ~ 0.28 | 0.028 ~ 0.28 |
| Fish lipid content (%) | 0.81 | 0.57 | 1.25 |
| BCF | <100 | <100 | <100 |
| Average BCF | | <100 | 1 |

The Shen and Hu (2008) study has been reviewed and was considered acceptable (see Appendix 1). Lack of detection in fish extracts (<0.028 mg/L) suggests a limited solubility in lipids and/or limited potential to partition into fish tissue from aqueous systems. However, there is some uncertainty associated with limit bounded values in any study because the absolute value is not known. But given the structure and likely behavior of disperse dyes in aqueous systems, a low BCF result is expected. Most disperse dyes, as their name suggests, exist as fine dispersible particles with limited truly soluble fractions. Solubility, however, can be increased by adding polar functional groups to the molecule. While AADM contains some of these solubilizing functional groups (nitro group), it is not predicted to solubilize at environmentally relevant pH (Table 2).

Therefore, given a melting point of 157° C (value for Disperse Blue 79 in Table 2) and a log K_{ow} of 4.45 (median of analogue data in Table 2), the predicted water solubility (WSKOWIN 2000) corrected for melting point and log K_{ow} is 0.075 mg/L which is within the aqueous detection limit in the bioaccumulation study and is in agreement with some of the analogue experimental solubility values for Disperse Blue 79 and 79:1 and Disperse Orange 30 (Table 2). Assuming that the concentration in solution in the test was equal to the water solubility value of 0.075 mg/L and using the fish concentration of 0.81 mg/kg as a worst case estimate, the BCF may be calculated to be <100.

While the above study serves as primary evidence to support AADM's lack of bioaccumulation potential, other research corroborate this conclusion. Anliker et al. (1981) reported experimental fish bioaccumulation values for 18 disperse monoazo dyes, performed according to test methods specified by the Japanese Ministry of International Trade and Industry (MITI). Expressed on the basis of wet body weight of the fishes, these log bioaccumulation factors ranged from 0.00 to 1.76 (Anliker et al. 1981). A lack of reporting of chemical registry numbers and chemical structures limited the utility of this study for read-across purposes to AADM. However, follow-up studies, which provided the chemical structures for the disperse dyes tested, confirmed low bioaccumulation potential for ten nitroazo dyes, with reported log bioaccumulation factors ranging from 0.3 to 1.76 (Anliker and Moser 1987; Anliker et al. 1988). Studies available from MITI also support low bioaccumulation potential for disperse azo dyes. Reported BCFs for three disperse azo dyes (CAS# 40690-89-9, 61968-52-3 and 71767-67-4) tested at a concentration of 0.01 mg/L were in the range of <0.3 to 47 (MITI 1992). An accumulation study by Brown (1987) also showed that none of the twelve disperse dyes tested accumulated during an eight week study with carp.

A high, median read-across log K_{ow} value of 4.45 for AADM's structural analogues (Table 2) is the only line of evidence that suggests AADM may have a high potential for bioaccumulation. In spite of the high K_{ow} values for AADM's structural analogues, evidence for bioaccumulation of disperse azo dyes is lacking (Anliker *et al.* 1981, Anliker and Moser 1987, Anliker *et al.* 1988, MITI 1992). Authors who have measured high log K_{ow} s and concomitant low bioaccumulation factors for disperse azo dyes suggest the low accumulation factors may be due in some cases to their low absolute fat solubility (Brown 1987) or their relatively high molecular weight (typically 450-550) which may make transport across fish membranes difficult (Anliker *et al.* 1981, Anliker and Moser 1987). It is also likely that the lack of bioavailability and limited capacity to partition under BCF test conditions limits accumulation in fish lipids.

It has been stated by ETAD (1995) that the molecular characteristics indicating the absence of bioaccumulation are a molecular weight of >450 g/mol and a cross-sectional diameter of >1.05 nm. Recent investigation by Dimitrov *et al.* (2002), Dimitrov *et al.* (2005) and the BBM (2008) suggests that the probability of a molecule crossing cell membranes as a result of passive diffusion declines significantly with increasing maximum cross-sectional diameter (D_{max}). The probability of passive diffusion lowers appreciably when cross-sectional diameter is > ~1.5 nm and more significantly for molecules having a cross-sectional diameter of >1.7 nm. Sakuratani *et al.* (2008) have

also investigated the effect of cross-sectional diameter on passive diffusion from a test set of about 1 200 new and existing chemicals, also observing that substances not having a very highly bioconcentration potential often have a $D_{max} > 2.0$ nm and an effective diameter (D_{eff}) >1.1 nm.

AADM has a molecular weight of 550.53 g/mol (see Table 1) and its molecular structure is relatively uncomplicated; the latter characteristic indicates a potential bioaccumulation capability. An Environment Canada (2007) report points out that there are no clear relationships for establishing strict molecular size cut-offs for assessing bioaccumulation potential. However, the report does not dispute the notion that a reduction in uptake rate can be associated with increasing cross-sectional diameter as demonstrated by Dimitrov *et al.* (2002, 2005). The maximum diameter of AADM and its conformers ranges from 1.56 to 2.14nm (BBM 2008) suggesting that a potential for a significantly reduced uptake rate from water and in vivo bioavailability exists with this dye.

Results of bioaccumulation modelling were not used in this assessment of AADM. Many higher molecular weight pigments and non-soluble dye classes, including disperse azo dyes are considered difficult to model and thus the results are generally unreliable. Predicted and/or empirical properties of disperse dyes relating to bioaccumulation (e.g., log K_{ow}) can be of uncertain relevance, or associated with a high degree of error that would limit the utility of calculated BCF and BAF values. In addition, disperse azo dyes fall outside of the domains of applicability of available bioaccumulation models.

Based on a lack of accumulation in bioconcentration tests of Disperse Orange 30 and other related disperse azo dyes, and AADM's large molecular size, which likely limits its partitioning behavior, AADM is expected to have a low potential for bioaccumulation. Therefore, taking into account analogue BCF evidence, and structural and bioavailability considerations, AADM does not meet the bioaccumulation criteria (BCF or BAF \geq 5000) as set out in the *Persistence and Bioaccumulation Regulations* (Canada 2000).

Potential to Cause Ecological Harm

Ecological Effects Assessment

A - In the Aquatic Compartment

No empirical ecotoxicity data were identified for AADM.

Environment Canada received ecotoxicological data for a structurally similar substance through the New Substance Notification Regulations (Environment Canada 1995). The results for the 96-hr static toxicity test with rainbow trout (*Oncorhynchus mykiss*) revealed that the LC₅₀ for this species is 505 mg/L (Table 7). The test was conducted according to OECD guideline No. 203. The Material Safety Data Sheets (MSDS) of the

notified substance also contained information on bacterial toxic effects. The results indicate activated sludge respiration inhibition $EC_{50} > 1\,000\,\text{mg/L}$. Based on the available ecotoxicity information, the notified substance is expected to be of low concern for toxic effects to aquatic organisms. Reliability of the study was assessed using a robust study summary and is considered to be satisfactory (Appendix 1).

In another study, a summary of which was submitted to Environment Canada on behalf of ETAD (Brown 1992), 11 disperse dyes were tested on the following organisms: zebra fish, Daphnia magna, algae and bacteria. Five of the dyes are azo analogues of AADM (Brown 1992). These are Disperse Red 73, Disperse Blue 79, Disperse Orange 25, Disperse Orange 30, and Disperse Red 17 (Table 7). The experimental details for the dyes tested were not provided, greatly limiting evaluation of these studies (Brown 1992). However, these data were considered usable and are included in this Screening Assessment as part of the available weight of evidence as they provide further empirical information to establish the range of ecotoxicity values for these structures. Two of the analogues tested show moderate toxicity to D. magna (48 hr EC₅₀=4.5-5.8 mg/L) and the five analogues showed moderate to low toxicity to zebra fish (96-hr LC₅₀=17 to 710 mg/L). Moderate toxicity was also presented for algae growth (EC₅₀ for growth = 6.7-54 mg/L) and no toxicity detected for bacteria (IC₅₀>100 mg/L). Lastly, an analogue, Disperse Blue 79:1, had a chronic 122-day no effect concentration (NOEC) for rainbow trout of > 0.0048mg/L (Table 7). Reliability of this study was assessed as high (Appendix 1). However, this value was not used to calculate the prediction no effect concentration because the value is a hypothesis-based unbounded result. These analogue values would also therefore suggest that AADM is not highly hazardous to aquatic organisms (i.e., acute LC_{50} are >1 mg/L).

Table 7. Empirical data for aquatic toxicity of AADM analogues

| Common Name | Test Organism | End point | Value (mg/L) | Reference |
|------------------|----------------------------|-------------------------------|--------------|------------|
| Disperse Orange | Zebra fish | LC_{50}^{-1} | 710 | Brown 1992 |
| 30 | Daphnia magna | EC_{50}^{2} | 5.8 | |
| | Scenedesmus subspicatus | $\mathrm{EC_{50}}^2$ | 6.7 | |
| | Bacteria | IC ₅₀ ³ | >100 | |
| Disperse Red 73 | Zebra fish | LC_{50}^{-1} | 17 | Brown 1992 |
| | Daphnia magna | EC_{50}^{2} | 23 | |
| | Scenedesmus | $\mathrm{EC_{50}}^2$ | >10 | |
| | subspicatus | | | |
| | Bacteria | IC_{50}^{3} | >100 | |
| Disperse Blue 79 | Zebra fish | LC ₅₀ ¹ | 340 | Brown 1992 |
| | Daphnia magna | EC_{50}^{2} | 4.5 | |
| | Scenedesmus subspicatus | EC ₅₀ ² | 9.5 | |
| | Bacteria | IC ₅₀ ³ | >100 | |
| Disperse Red 17 | Zebra fish | IC_{50}^{3} | 103 | Brown 1992 |
| | Daphnia magna | LC ₅₀ ¹ | 98 | |
| | Scenedesmus subspicatus | EC ₅₀ ² | 7 | |

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| | Bacteria | $\mathrm{EC_{50}}^2$ | >100 | |
|---------------------------|----------------------------|-------------------------------|--------|----------------------------|
| Disperse Orange | Zebra fish | IC_{50}^{3} | 268 | Brown 1992 |
| 25 | Daphnia magna | LC ₅₀ ¹ | 110 | |
| | Scenedesmus subspicatus | EC_{50}^{2} | 54 | |
| | Bacteria | $\mathrm{EC_{50}}^2$ | >100 | |
| Analogue disperse azo dye | Rainbow trout | LC_{50}^{-1} | 505 | Environment Canada 1995 |
| Disperse Blue 79:1 | Rainbow trout | NOEC ⁴ (122 days) | 0.0048 | Cohle and Mihalik 1991 |
| Disperse Yellow 3 | Fathead minnow | LC_{50}^{-1} | >180 | Little and Lamb 1973 |

 $^{^{1}}$ LC₅₀ – The median concentration of a substance that is estimated to be lethal to 50% of the test organisms.

In general, due to their low solubility (<1 mg/L) disperse dyes are expected to have a low acute ecological impact (Hunger 2003). The results of empirical toxicity studies with several analogues of AADM are consistent with this expectation, indicating LC₅₀s in the 5 to 710mg/L range, with *Daphnia* being the most sensitive organism tested (EC₅₀/LC₅₀s from 4.5 to >100 mg/L). Although interpretation of results from these tests is complicated by the fact that the reported effect values (*i.e.* EC₅₀ and LC₅₀s) are likely to be much greater than the solubility of the substances tested as well as AADM, the analogue data available do indicate that the toxicity of AADM is likely to be low.

A range of aquatic toxicity predictions for AADM were also obtained from the various QSAR models considered for AADM and its analogues. However, as with bioaccumulation, these QSAR ecotoxicity predictions for AADM are not considered reliable because of the unique nature of disperse dyes, specifically structural and/or physico-chemical properties which fall outside of the models' domain of applicability.

The available empirical ecotoxicity information for analogues of AADM thus indicates that it is not likely to be highly hazardous to aquatic organisms.

B - In Other Environmental Compartments

Since AADM may potentially enter soil from biosludge which is commonly used for soil enrichment as well as from the disposal of products that degrade and release AADM, it would be desirable to obtain toxicity data for soil organisms. However, no suitable ecological effects studies were found for this compound or analogues in media other than water. The toxicity potential is also likely to be low in sediment dwelling species considering the lack of bioaccumulation potential and bioavailability as well as the

 $^{^{2}}$ EC₅₀ – The median concentration of a substance that is estimated to cause some toxic sublethal effect on 50% of the test organisms.

 $^{^{3}}$ IC $_{50}$ – The median concentration of a substance that is estimated to cause inhibition to growth 50% of the test organisms.

⁴NOEC – The concentration at which no effects have been observed.

physical-chemical makeup of AADM, although this cannot be substantiated due to lack of whole organism sediment toxicity data for AADM or suitable analogues.

Ecological Exposure Assessment

No data concerning concentrations of this substance in water in Canada have been identified. Environmental concentrations are, therefore, estimated from available information, including estimated substance quantities, release rates, and receiving water bodies.

The Mass Flow Tool identified releases to the water (sewer) from formulation use and from consumer use of products containing this substance (Table 4). To address releases from industrial activities, Environment Canada's Industrial Generic Exposure Tool – Aquatic (IGETA) was employed to estimate the substance concentration (worst-case) in a generic water course receiving industrial effluents (Environment Canada 2008c). The generic scenario is designed to provide these estimates based on conservative assumptions regarding the amount of chemical processed and released, the number of processing days, (STP) treatment plant removal rate, and the size of the receiving watercourse. The tool models an industrial-release scenario based on loading data from sources such as industrial surveys and knowledge of the distribution of industrial discharges in the country, and calculates a predicted environmental concentration (PEC). The equation and inputs used to calculate the PEC in the receiving water course are described in the Environment Canada (2008d). The amount of AADM used was conservatively assumed to be 100 kg, the S.71 reporting threshold. As a conservative estimate, the release to water (sewer) from industrial activities only was set at 16% of the amount used as per previous experience of Environment Canada on assessing other disperse azo dyes. Conservative assumptions were made regarding receiving water body, by assuming the chemical is released to a very small river with no removal from sewage treatment plants. The conservative PEC for water was calculated to be 0.0018 mg/L (Environment Canada 2008d).

To address down-the-drain releases from consumer uses, Environment Canada's spreadsheet model (Mega Flush) was used. Using Mega Flush, potential substance concentrations in multiple water bodies receiving sewage treatment plant effluents to which consumer products containing the substance may have been released is performed (Environment Canada 2008e). The spreadsheet model is designed to provide these estimates based on conservative assumptions regarding the amount of chemical used and released by consumers. By default, primary and secondary STP removal rates are assumed to be of 0%, fraction release during use of 100%, consumer use of the substance is over 365 days/year, and the flow rate used for receiving water bodies at all sites is the 10th percentile value. These estimates are made for approximately 1000 release sites across Canada, which account for most of the major STPs in Canada. These parameter values are considered to result in a very conservative scenario.

The equation and inputs used in Mega Flush to calculate the PEC of AADM in the receiving water bodies are described in Environment Canada (2008f). The predicted releases to water (sewers) from formulation use and from consumer use of products containing this substance were based on previous experience with disperse azo dyes. The consumer use quantity was estimated conservatively based on reporting threshold from the S. 71 survey (100 kg), and the ratio of textiles manufactured in Canada / imported textiles of 30/70. A 10% loss of dye was then assumed for the total amount of the substance being used by consumers (Øllgaard *et al.* 1998). Thus 28.1 kg of AADM were predicted to be released to water, as a result of loss to sewers during the laundering of manufactured articles that contain this dye but are manufactured in Canada. Primary and secondary STP removal rates of 0% were used. These assumptions result in a very conservative scenario. Using this scenario, the Mega Flush tool estimates that the PEC in the receiving water bodies ranges from 0.0000035 to 0.000043 mg/L.

Characterization of Ecological Risk

A predicted no-effect concentration (PNEC) was estimated based on the lowest nominal sublethal concentration (EC₅₀) to *D. magna* for an analogue of AADM. The 96-hour EC₅₀ for Disperse Blue 79 (CAS RN12239-34-8), an analogue of AADM, was 4.5 mg/L (Table 7). A factor of 100 was then applied to account for extrapolating from acute to chronic (long-term) toxicity and from laboratory results for one species to other potentially sensitive species in the field. The resulting PNEC is 0.045 mg/L.

When compared to the conservative PEC calculated above using IGETA, the resulting risk quotient for industrial discharges (PEC/PNEC) is 0.0018/0.045 = 0.04. Therefore, concentrations of AADM in surface waters in Canada appear unlikely to cause adverse effects to aquatic organisms. Given that IGETA provides a conservative estimate of exposure, the results indicate a low potential for ecological harm resulting from local exposure to point source industrial release.

For exposure resulting from down-the-drain releases through consumer uses (conservative scenario), Mega Flush results estimate that the PEC will not exceed the PNEC at any site (i.e., all risk quotients < 1). This indicates that down-the-drain consumer releases of AADM are not expected to harm aquatic organisms.

Based on the available information, AADM is expected to be persistent in water, soil and sediment; it is however expected to have a low bioaccumulation potential. The lack of reports of manufacture and the likely very low importation quantities of AADM into Canada, along with information on physical and chemical properties and its uses, indicate a low potential for releases into the Canadian environment. If released into the environment, it is expected that AADM will be mainly discharged to surface waters where ultimately it is expected to be transferred to sediment. It is also expected to have only a low to moderate potential for acute toxicity to aquatic organisms. Risk quotients for aquatic exposures indicate that AADM concentrations likely do not exceed

concentrations associated with effects, even when using conservative scenarios and assumptions. Therefore AADM is unlikely to be causing harm to populations of aquatic organisms in Canada.

Uncertainties in Evaluation of Ecological Risk

An area of uncertainty for AADM is associated with the use of read-across data for physical and chemical properties, as well as toxicity data from analogues. While the chemicals identified (Disperse Blue 79, Disperse Blue 79:1, Disperse Orange 30, Disperse Orange 25, Disperse Red 17, Disperse Red 73 and Disperse Yellow 3), share many similarities with AADM, including being azo dyes with high molecular weights, similar cross sectional diameters, having solid particulate structures that decompose at greater than 74 °C (to 240 °C), and being "dispersible" in water (i.e., not truly soluble), they do have some differences in functional groups. These differences in chemical structure add uncertainty because the properties and toxicity of AADM may be somewhat different. However, it was reasoned that the similarities were sufficient to include the data from analogues to contribute to the weight of evidence in the assessment of AADM.

The persistence assessment is limited by the absence of biodegradation data, which necessitated generation of model predictions. Although all model prediction has some degree of error, the aerobic biodegradation model outputs confirmed the expected persistence of AADM given its uses and structural characteristics. In addition, the persistence assessment is limited by the uncertainty about the rate and extent to which degradation occurs in anaerobic sediments and whether the degradation products (e.g., amines) would be biologically available. Nevertheless it is clear that anaerobic degradation of the bioavailable portion azo dyes in sediments to constitutive amines is much faster (half-lives in the order of days) than aerobic biodegradation. Although the amine degradation products are not expected to be biologically available because they form only in relatively deep anoxic sediment and can be irreversibly bound to sediment through nucleophilic addition and oxidative radical coupling (Colon *et al.* 2002, Weber *et al.* 2001), this issue is a source of uncertainty in the assessment of AADM.

Uncertainties are present due to the lack of bioaccumulation studies for this substance. However, based on a lack of accumulation in bioconcentration tests in Disperse Orange 30 and other related disperse azo dyes and AADM large molecular size, which likely limits its partitioning behavior, AADM's is expected to have a low potential for bioaccumulation.

Uncertainties are also present due to the lack of information on environmental concentrations in Canada for AADM. However, the lack of reports of manufacturing in or import of the chemical into Canada suggests low release of this chemical into the Canadian environment.

The experimental concentrations associated with toxicity for aquatic organisms may have an additional source of uncertainty when these concentrations exceed the solubility of the

chemical in water (either experimental or predicted). Despite this, the available data indicate that AADM is not highly hazardous to aquatic organisms.

Uncertainties are also associated with the fraction of the substance that is released during use. These uncertainties were addressed by making conservative assumptions using best model estimates.

Regarding ecotoxicity, based on the predicted partitioning behaviour of this chemical, the significance of soil and sediment as important media of exposure is not well addressed by the effects data available. Indeed, the only effects data identified apply primarily to pelagic aquatic exposures.

Conclusion

Based on the information presented in this screening assessment, it is concluded that AADM is not entering the environment in a quantity or concentration or under conditions that have or may have an immediate or long-term harmful effect on the environment or its biological diversity or that constitute or may constitute a danger to the environment on which life depends.

It is therefore concluded that AADM does not meet the definition of toxic as set out in section 64 of CEPA 1999. Additionally, AADM meets the criteria for persistence but does not meet the criteria for bioaccumulation potential as set out in the *Persistence and Bioaccumulation Regulations* (Canada 2000).

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Appendix I - Robust Study Summaries for Key Studies

| | Robust Study Summaries Form: Aquatic B | | | | | | | | | |
|---------------|---|--------|----------|---|--|--|--|--|--|--|
| No | Item | Weight | Yes/No | Specify | | | | | | |
| 1 | hen, Genxiang and Hu, Shuangqing. 2008. Bioconcentration Test of C.I. Disperse Orange 30 in Fish. repared by Environmental Testing Laboratory, Shanghai Academy of Environmental Sciences, Shanghai, hina for Dystar in the name of Ecological and Toxicological Association of the Dyes and Organic igments Manufacturers (ETAD) Basel, Switzerland. Report No. S-070-2007. Submitted to Environment anada in April 2008. Challenge Submission ID#8351 | | | | | | | | | |
| 2 | Substance identity: CAS RN | n/a | Υ | 5261-31-4 | | | | | | |
| 3 | Substance identity: chemical name(s) | n/a | Y | Propanenitrile, 3-[[2-(acetyloxy)ethyl][4-[(2,6-dichloro-4-nitrophenyl)azo]phenyl]amino]- | | | | | | |
| 4 | Chemical composition of the substance | 2 | N | 7, 41 - 7, | | | | | | |
| 5 | Chemical purity | 1 | N | | | | | | | |
| 6 | Persistence/stability of test substance in aquatic solution reported? | 1 | N | | | | | | | |
| 7 | If test material is radiolabelled, were precise position(s) of the labelled atom(s) and the percentage of radioactivity associated with impurities reported? | 2 | n/a | | | | | | | |
| | Method | | | | | | | | | |
| 8 | Reference | 1 | Υ | OECD guidelines for the testing of chemicals No 305B-1996 | | | | | | |
| 9 | OECD, EU, national, or other standard method? | 3 | Y | OECD | | | | | | |
| 10 | Justification of the method/protocol if not a standard method was used | 2 | | | | | | | | |
| 11 | GLP (Good Laboratory Practice) | 3 | N | | | | | | | |
| Test organism | Test organism | | | | | | | | | |
| 12 | Organism identity: name | n/a | <u>Y</u> | zebra fish, Brachydanio rerio | | | | | | |
| 13 | Latin or both Latin & common names reported? | 1 | Y | both | | | | | | |
| 14 | Life cycle age / stage of test organism | 1 | N | | | | | | | |
| 15 | Length and/or weight | 1 | Y | Mean body length 3.91+/-0.18cm and mean body weight 0.32+/-0.06g | | | | | | |
| 16 | Sex | 1 | N | | | | | | | |
| 17 | Number of organisms per replicate | 1 | Υ | 7 | | | | | | |
| 18 | Organism loading rate | 1 | Υ | 20mg/L | | | | | | |
| 19 | Food type and feeding periods during the acclimation period | 1 | Y | Fed a commercial fish diet until one day before start of test | | | | | | |
| | Test design / conditions | | | | | | | | | |
| 20 | Experiment type (laboratory or field) | n/a | Y | Laboratory | | | | | | |
| 21 | Exposure pathways (food, water, both) | n/a | Y | Water | | | | | | |
| 22 | Exposure duration | n/a | Y | 28 days | | | | | | |
| 23 | Number of replicates (including controls) | 1 | Y | | | | | | | |
| 24 | Concentrations | 1 | Y | 20 mg/L | | | | | | |
| 25 | Food type/composition and feeding periods during the test | 1 | Y | Fish were fed two hours before water renewal | | | | | | |
| 26 | If BCF/BAF derived as a ratio of chemical concentration in the organism and in water, was experiment duration equal to or longer than the time required for the chemical concentrations to reach steady state? | 3 | Y | 28 days | | | | | | |

| 50 | Comments | every 2 also be | present procedure is based on semi-static conditions (renewal of test solutions y 2 days). Therefore, test chemical with very low water solubility like AADM, can be characterized as to their bioconcentration potential without adding solvents or rauxiliary substances which may affect the results. | | | | |
|----|--|------------------------|--|-----|---|--|--|
| 49 | Reliability category (high, satisfactory, low): | | Satisfactory Confidence | | | | |
| 48 | EC Reliability code: | | 2 | | | | |
| 47 | Score: % | | 75.0 | | | | |
| 46 | Whether 1) average or 2) maximum BAF/BCF was used? | 1 | n/a n/a 1 | | | | |
| 45 | tissue sample or 2) whole organism | า? | n/a | n/a | 2 | | |
| 44 | BAF or BCF determined as: 1) the inchemical concentration in the orgal and in water, or 2) the ratio of the cuptake and elimination rate constant. Whether BAF/BCF was derived from | nism hemical nts | n/a | n/a | 1 | | |
| 43 | Endpoints (BAF, BCF) and values | ratic of | n/a | n/a | BCF < 100 | | |
| 40 | Results | | - 1 | - 1 | DOE 100 | | |
| 42 | was BCF determination based on the parent compound (i.e. not on total radiolabelled residues)? | | 3 | n/a | | | |
| 41 | Were measured concentrations of a chemical in the test water below the chemical's water solubility? If radiolabelled test substance was | 9 | 3 | N | | | |
| 40 | Was lipid content (or lipid-normalize BAF/BCF) reported? | | 2 | Y | | | |
| 39 | Was temperature of the test water water the range typical for the Canadian environment (5 to 27°C)? | | 1 | Υ | 22-23 | | |
| 38 | Was pH of the test water within the range typical for the Canadian environment (6 to 9)? | | 1 | Υ | 7.22-7.84 | | |
| 37 | Does system type and design (static, semi- static, flow-through; sealed or open; etc.) correspond to the substance's properties and organism's nature/habits? | | 2 | Υ | Semi-static | | |
| 36 | Were the test conditions (pH, temporal DO, etc.) typical for the test organisms. | sm? | 1 | Y | | | |
| 35 | Was the test organism relevant to the Canadian environment? | | 3 | Y | | | |
| | Information relevant to the data | | | | | | |
| 34 | chemical was unstable or poorly so | luble? | n/a | N | | | |
| 33 | Statistical methods used Was solubilizer/emulsifier used, if the | he | 1 | Y | | | |
| 32 | Analytical monitoring intervals | | 1 | Y | temperature | | |
| 31 | Stock and test solution preparation | | 1 | Y | Every second day for dissolved oxygen, pH and | | |
| 30 | Photoperiod and light intensity | | 1 | Y | 12:12 | | |
| 29 | Were the exposure media conditions relevant to the particular chemical reported? (e.g., for the metal toxicity - ph DOC/TOC, water hardness, temperature | | 3 | Y | Yes every second day | | |
| 28 | Were the exposure media condition | | 1 | Y | On three separate days | | |
| 27 | If BCF/BAF derived as a ratio of checoncentration in the organism and i were measured concentrations in b water and organism reported? | in water, ooth | 3 | Y | | | |

| | Robust Study Summaries Form: Aquatic iT | | | | | |
|---|--|--------|--------|---|--|--|
| No | Item | Weight | Yes/No | Specify | | |
| Reference: Environment Canada. 1995. Acute fish toxicity test submission in fulfillment of new substances notification regulations to New Substances Branch, Environment Canada under New Substance Notification Program. | | | | | | |
| 2 | Substance identity: CAS RN | n/a | N | | | |
| 3 | Substance identity: chemical name(s) | n/a | Υ | | | |
| 4 | Chemical composition of the substance | 2 | N | | | |
| 5 | Chemical purity | 1 | N | | | |
| 6 | Persistence/stability of test substance in aquatic solution reported? | 1 | N | | | |
| | Method | • | • | | | |
| 7 | Reference | 1 | Υ | OECD 203 | | |
| 8 | OECD, EU, national, or other standard method? | 3 | Υ | | | |
| 9 | Justification of the method/protocol if not a standard method was used | 2 | | not applicable | | |
| 10 | GLP (Good Laboratory Practice) | 3 | Y | | | |
| | Test organism | 1 | L | | | |
| 11 | Organism identity: name | n/a | Y | Rainbow trout | | |
| 12 | Latin or both Latin & common names reported? | 1 | Υ | | | |
| 13 | Life cycle age / stage of test organis | 1 | Y | mean length 51mm and mean weight 1.54 | | |
| 14 | Length and/or weight | 1 | Y | see above | | |
| 15 | Sex | 1 | | not applicable | | |
| | Number of organisms per replicate | 1 | Y | 10 | | |

| | | Ī | i | | | | | |
|----|--|------------|---|------------------|--|--|--|--|
| 17 | Organism loading rate | 1 | Y | | | | | |
| 18 | Food type and feeding periods during the acclimation period | 1 | Υ | | | | | |
| | Test design / conditions | | | | | | | |
| 19 | Test type (acute or chronic | n/a | Y | acute | | | | |
| 20 | Experiment type (laboratory or field | n/a | У | lab | | | | |
| 21 | Exposure pathways (food, water, both) | n/a | У | water | | | | |
| 22 | Exposure duration | n/a | У | 96hrs | | | | |
| 23 | Negative or positive controls (specify) | 1 | Y | 3 | | | | |
| 24 | Number of replicates (including controls) | 1 | Y | 2 | | | | |
| 25 | Nominal concentrations reported? | 1 | Y | 320 to 3200 mg/L | | | | |
| 26 | Measured concentrations reported? | 3 | N | 020 to 0200 mg/2 | | | | |
| | • | | | | | | | |
| 27 | Food type and feeding periods during the long-term tests | 1 | | not applicable | | | | |
| 28 | Were concentrations measured periodically (especially in the chronic test)? | 1 | N | | | | | |
| 29 | Were the exposure media conditions relevant to the particular chemical reported? (e.g., for the metal toxicity - pH, DOC/TOC, water hardness, temperature) | 3 | Y | | | | | |
| 30 | Photoperiod and light intensity | 1 | Υ | | | | | |
| 31 | Stock and test solution preparation | 1 | Y | | | | | |
| 32 | Was solubilizer/emulsifier used, if the chemical was poorly soluble or unstable? | 1 | N | | | | | |
| 33 | If solubilizer/emulsifier was used, was its concentration reported? | 1 | | | | | | |
| 34 | If solubilizer/emulsifier was used, was its ecotoxicity reported? | 1 | | | | | | |
| 35 | Analytical monitoring intervals | 1 | Y | | | | | |
| 36 | Statistical methods used | 1 | Υ | | | | | |
| | Information relevant to the d | ata qualit | У | | | | | |
| 37 | Was the endpoint directly caused by the chemical's toxicity, not by organism's health (e.g. when mortality in the control >10%) or physical effects (e.g. 'shading effect')? | n/a | Y | | | | | |
| 38 | Was the test organism relevant to the Canadian environment? | 3 | Υ | | | | | |
| 39 | Were the test conditions (pH, temperature, DO, etc.) typical for the test organism? | 1 | Υ | | | | | |
| 40 | Does system type and design (static, semi-static, flow-through; sealed or open; etc.) correspond to the substance's properties and organism's nature/habits? | 2 | Y | | | | | |
| 41 | Was pH of the test water within the range typical for the Canadian environment (6 to 9)? | 1 | Υ | | | | | |

| 42 | Was temperature of the test water within the range typical for the Canadian environment (5 to 27°C)? | 1 | Y | | | |
|----|--|-------------------------|-----|-----------------------------|--|--|
| 43 | Was toxicity value below the chemical's water solubility? | 3 | | unknown water solubility | | |
| | Results | | | | | |
| 44 | Toxicity values (specify endpoint and value) | n/a | n/a | 96hr LC50=505 mg/L | | |
| 45 | Other endpoints reported - e.g. BCF/BAF, LOEC/NOEC (specify)? | n/a | N | | | |
| 46 | Other adverse effects (e.g. carcinogenicity, mutagenicity) reported? | n/a | N | | | |
| 47 | 77.5 Score: % | | | | | |
| 48 | 48 EC Reliability code: 2 | | | | | |
| 49 | Reliability category (high, satisfactory, low): | Satisfactory Confidence | | | | |
| 50 | Comments | | | | | |

| | Robust Study Summaries Form: Aquatic iT | | | | | |
|--|--|----------|--------|--------------------|--|--|
| No | Item | Weight | Yes/No | Specify | | |
| Reference: Cohle P, R Mihalik R. 1991. Early life stage toxicity of C.I. Disperse Blue 79:1 purified preecake to rainbow trout (<i>Oncorhynchus mykiss</i>) in a flow-through system. Final report. ABC Laboratories Inc. Columbia MO. | | | | | | |
| 2 | Substance identity: CAS RN | n/a | | | | |
| 3 | Substance identity: chemical name(s) | n/a | | Disperse Blue 79:1 | | |
| 4 | Chemical composition of the substance | 2 | | n/a | | |
| 5 | Chemical purity | 1 | Y | 96.61 | | |
| 6 | Persistence/stability of test substance in aquatic solution reported? | 1 | N | | | |
| | Method | | | | | |
| 7 | Reference | 1 | Υ | | | |
| 8 | OECD, EU, national, or other standard method? | 3 | Υ | | | |
| 9 | Justification of the method/protocol if not a standard method was used | 2 | | n/a | | |
| 10 | GLP (Good Laboratory Practice) | 3 | Y | | | |
| | Test organism | <u> </u> | | | | |
| 11 | Organism identity: name | n/a | | Rainbow trout | | |
| 12 | Latin or both Latin & common names reported? | 1 | Υ | | | |
| 13 | Life cycle age / stage of test organis | 1 | Y | | | |
| 14 | Length and/or weight | 1 | Y | | | |
| 15 | Sex | 1 | | n/a | | |
| 16 | Number of organisms per replicate | 1 | Υ | 20 | | |
| 17 | Organism loading rate | 1 | Y | 0.36 to 4.8ug/L | | |

| 18 | Food type and feeding periods during the acclimation period | 1 | Y | |
|----|--|------------|----|---|
| | Test design / condition | ons | • | |
| 19 | Test type (acute or chronic | n/a | Y | chronic |
| 20 | Experiment type (laboratory or field | n/a | Υ | lab |
| 21 | Exposure pathways (food, water, both) | n/a | Υ | water |
| 22 | Exposure duration | n/a | Y | 122 days |
| 23 | Negative or positive controls (specify) | 1 | Y | control and carrier blank |
| 24 | Number of replicates (including controls) | 1 | Υ | 2 |
| 25 | Nominal concentrations reported? | 1 | Y | 5 |
| 26 | Measured concentrations reported? | 3 | Y | Ů |
| 27 | Food type and feeding periods during the long-term tests | 1 | Y | |
| 28 | Were concentrations measured periodically (especially in the chronic test)? | 1 | Y | |
| 29 | Were the exposure media conditions relevant to the particular chemical reported? (e.g., for the metal toxicity - pH, DOC/TOC, water hardness, temperature) | 3 | Y | |
| 30 | Photoperiod and light intensity | 1 | Υ | |
| 31 | Stock and test solution preparation | 1 | Υ | |
| 32 | Was solubilizer/emulsifier used, if the chemical was poorly soluble or unstable? | 1 | Υ | |
| 33 | If solubilizer/emulsifier was used, was its concentration reported? | 1 | Υ | |
| 34 | If solubilizer/emulsifier was used, was its ecotoxicity reported? | 1 | Υ | no tox value but however is was used as a control |
| 35 | Analytical monitoring intervals | 1 | Υ | |
| 36 | Statistical methods used | 1 | Υ | |
| | Information relevant to the d | ata qualit | 'V | • |
| 37 | Was the endpoint directly caused by the chemical's toxicity, not by organism's health (e.g. when mortality in the control >10%) or physical effects (e.g. 'shading effect')? | n/a | Y | |
| 38 | Was the test organism relevant to the Canadian environment? | 3 | Y | |
| 39 | Were the test conditions (pH, temperature, DO, etc.) typical for the test organism? | 1 | Y | |
| 40 | Does system type and design (static, semi-static, flow-through; sealed or open; etc.) correspond to the substance's properties and organism's nature/habits? | 2 | Y | flow through |
| 41 | Was pH of the test water within the range typical for the Canadian environment (6 to 9)? | 1 | Y | |
| 42 | Was temperature of the test water within the range typical for the Canadian environment (5 to 27°C)? | 1 | Y | |

| 43 | Was toxicity value below the chemical's water solubility? | 3 | | n/a |
|----|--|-----|-----|------------|
| | Results | | | |
| 44 | Toxicity values (specify endpoint and value) | n/a | n/a | NOEC>5ug/L |
| 45 | Other endpoints reported - e.g. BCF/BAF, LOEC/NOEC (specify)? | n/a | | |
| 46 | Other adverse effects (e.g. carcinogenicity, mutagenicity) reported? | n/a | | |
| 47 | Score: % | | 9 | 7.6 |
| 48 | EC Reliability code: | | | 1 |
| 49 | 49 Reliability category (high, satisfactory, low): | | | onfidence |
| 50 | Comments | | | |