

CPHLN recommendations for the laboratory detection of Shiga toxin-producing *Escherichia coli* (O157 and non-O157)

L Chui^{1*}, S Christianson², DC Alexander³, V Arseneau⁴, S Bekal⁵, B Berenger¹, Y Chen⁶, R Davidson⁷, DJ Farrell⁸, GJ German⁴, L Gilbert⁹, LMN Hoang¹⁰, RP Johnson², A MacKeen¹¹, A Maki⁶, C Nadon², E Nickerson¹², A Peralta⁶, SM Radons Arneson¹¹, Y Yu⁶, K Ziebell² on behalf of Canadian Public Health Laboratory Network (CPHLN)

Abstract

Shiga toxin-producing *Escherichia coli* (STEC) are important enteric pathogens responsible for sporadic cases and outbreaks of gastroenteritis. *E.coli* O157:H7/NM (STEC O157) are the most commonly known STEC serotypes but it is now increasingly apparent that non-O157 STEC serotypes have been underreported in the past because they were not part of routine screening in many front-line laboratories. The Canadian Public Health Laboratory Network (CPHLN) has identified the need for improved detection and surveillance of non-O157 STEC and has developed the following recommendations to assist in the decision-making process for clinical and reference microbiology laboratories. These recommendations should be followed to the best of a laboratory's abilities based on the availability of technology and resources.

The CPHLN recommends that when screening for the agents of bacterial gastroenteritis from a stool sample, front-line laboratories use either a chromogenic agar culture or a culture-independent diagnostic test (CIDT). CIDT options include nucleic acid amplification tests (NAATs) to detect Shiga toxin genes or enzyme immunoassays (EIAs) to detect Shiga toxins. If either CIDT method is positive for possible STEC, laboratories must have a mechanism to culture and isolate STEC in order to support both provincial and national surveillance as well as outbreak investigations and response. These CPHLN recommendations should result in improved detection of STEC in patients presenting with diarrhea, especially when due to the non-O157 serotypes. These measures should enhance the overall quality of healthcare and food safety, and provide better protection of the public via improved surveillance and outbreak detection and response.

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Affiliations

- ¹ Provincial Laboratory for Public Health, Alberta Public Laboratories, AB
- ² National Microbiology Laboratory, MB & ON
- ³ Cadham Provincial Laboratory, MB
- ⁴ Health Prince Edward Island, PE
- ⁵ Laboratoire de Santé Publique du Québec, QC
- ⁶ Public Health Ontario, ON
- ⁷ Queen Elizabeth II Health Science Centre, NS
- ⁸ Roy Romanow Provincial Laboratory, SK
- ⁹ Provincial Public Health and Microbiology Laboratory, Eastern Health, NL
- ¹⁰ BC Centre for Disease Control Public Health Laboratory, BC
- ¹¹ Canadian Public Health Laboratory Network, MB
- ¹² The New Brunswick Enteric Reference Centre, NB

*Correspondence:

linda.chui@ahs.ca

Introduction

Escherichia coli are part of the normal flora of the gut. However, Shiga toxin-producing Escherichia coli (STEC) are intestinal pathogens. Although they typically cause a self-limited episode of diarrhea and abdominal pain, on rare occasions they cause severe—and potentially fatal—sequelae such as hemolytic uremic syndrome (1).

E. coli O157:H7/NM (STEC O157) are the most common STEC serotypes causing infection in humans, but many non-O157 STEC serotypes have been associated with serious illness and major outbreaks (2). In 2011 there was a European outbreak of E. coli O104:H4 (3). In 2016, there was an outbreak in Canada and the United States of STEC O121 infections associated with flour (4).



These recent outbreaks of non-O157 serotypes have highlighted the clinical importance of timely and reliable detection and surveillance of these organisms (5–10).

In 2016, 642 cases of STEC infections were reported to the Canadian National Enteric Surveillance Program (NESP); approximately 35% were caused by non-O157 STEC (11). Current data for Canada on non-O157 STEC infections are likely an underestimate as they are not part of routine screening in many laboratories (11,12). There are several reasons for this gap. STEC O157 is readily identified using differential and selective media such as Sorbitol-MacConkey agar and chromogenic O157 agar. Unlike O157 STEC, non-O157 STEC lack the phenotypic characteristics that readily distinguish them from generic *E. coli*, making culture-based isolation challenging.

Detection of non-O157 STEC subtypes

Improvements to laboratories' ability to identify non-O157 STEC are based on the use of chromogenic and/or selective agars and culture-independent diagnostic tests (CIDT). The two common CIDTs include nucleic acid amplification tests (NAATs) for detection of Shiga toxin genes, and enzyme immunoassays (EIAs) that detect Shiga toxins. Less common CIDTs include Shiga toxin detection by cell culture cytotoxicity assays or reverse passive latex agglutination test, and isolation of selected serogroups by O-antigen immunomagnetic bead-capture methods. However, these methods are not practical in most front-line microbiology laboratories. Not only is it important to identify the STEC subtype, it is also essential to have isolates for further characterization such as serotyping, molecular genotyping and whole genome sequencing.

Each detection method has its advantages and disadvantages. Chromogenic agar has a sensitivity of greater than 85% (13). It is less costly than other methods and can be easily substituted into a laboratory's workflow by replacing the current O157specific plates. However, chromogenic agar can be inhibitory to some STEC serotypes (14). As for the detection of Shiga toxins by EIA, both microwell and lateral flow formats are available. The sensitivity of EIAs is lower for direct stool testing; however, overnight enrichment in MacConkey broth (15) or other suitable broth may provide sensitivity approaching that of NAATs (13,16). Both in-house and commercial NAATs have equivalent sensitivities and are the most sensitive methods available (15,17). Many NAATs can be performed directly on stool samples (18), improving the turn-around time compared to culture and EIA. Multiplex NAATs have the added advantage of detecting multiple pathogens concurrently. Both EIAs and NAATs will detect Shiga toxins and Shiga toxin genes respectively from any serotype of STEC. However, EIAs and NAATs are more expensive than agar for screening. NAATs also require laboratories to purchase additional equipment, designated space for molecular set-up and specialized training of personnel, which may be impractical for many frontline laboratories.

Public health implications

Isolation and further subtyping enables the comparison of STEC strains in order to identify outbreaks and potential sources of infection. Once the suspected source of the outbreak is identified, tracing and outbreak management activities by public health officials can prevent further transmission and promote public awareness.

The objectives of the following recommendations are to identify laboratory choices in a testing workflow for the detection, confirmation of STEC in stool specimens, and recovery of positive isolates for further characterization.

Recommendations

The Canadian Public Health Laboratory Network (CPHLN) recommends that when screening for the agents of bacterial gastroenteritis, front-line laboratories use one or more of three options for the detection of STEC: NAAT, culture on selective agar, or broth enrichment plus an EIA (**Figure 1**). Stool specimens submitted for STEC detection should follow local guidelines for submission and testing. Laboratories using chromogenic agar as their primary screening method may consider using an EIA method with broth enrichment or a NAAT in selected cases where there is a high suspicion of STEC infection and chromogenic agar results are negative.

If CIDT is implemented for STEC testing, culture is still recommended for the recovery of isolates for further characterization when Shiga toxins or Shiga toxin genes are identified. CPHLN culture recommendations following CIDT can be found in Berenger et al. (19). It is imperative that front-line laboratories communicate with their referral public health laboratories to determine the required work-up for culture, isolation or characterization of isolates before submitting any samples.

Discussion

To accurately diagnose all STEC-related cases of gastroenteritis that may lead to outbreaks and have public health implications, it is important that both O157 and non-O157STEC serotypes are identified. To facilitate this, screening should be done using either culture or CIDT by a NAAT or an EIA. If the CIDT is positive for possible STEC, then culture is needed for STEC confirmation and characterization.

Laboratories should follow the CPHLN recommendations to the best of their abilities based on the availability of technology and resources. It is important to emphasize that when STEC are detected, culturing the organism is of paramount importance for further characterization (typing and subtyping), as well as



Stool (collected and submitted as per local protocols) **Immunoassay Nucleic Acid Tests** Immunoassav Nucleic Acid (Overnight enrichment in Test for stx MacConkey or other suitable broth) Culture Positive Negative Report Report: No stx detected stx detected Report: Report: Stx detected No Stx detected culture on STEC selective Follow Provincial Follow Provincial and differential Procedures for Procedures for agar Culture Culture (e.g. chromogenic) Submissions Submissions Negative for STEC Positive Report: No STEC isolated (for possible STEC) LEGEND STEC Detection, CPHLN: Canadian Public Health Laboratory

Confirmation and

Characterization

as per laboratory protocols

Figure 1: Recommendations for the detection of Shiga toxin-producing Escherichia coli in stool specimens

to facilitate the outbreak response and support surveillance programs, including PulseNet Canada and the National Enteric Surveillance Program.

Network

stx: Shiga toxin gene Stx: Shiga toxins

STEC: shiga toxin-producing E. coli

The role of the provincial or federal laboratories is to support front-line laboratories by performing confirmatory testing as necessary, as well as serotyping and other reference laboratory services for STEC isolates. These laboratories are also available to assist with facilitating the implementation of these new testing algorithms for non-O157 STEC. The public health laboratory system varies among provinces; so any changes in public health laboratory protocols that may impact the capacity of front-line laboratories to follow these recommendations must be discussed with the front-line laboratories and other stakeholders prior to implementation.

Conclusion

Following these CPHLN recommendations should result in improved detection of STEC in patients presenting with diarrhea, especially the non-O157 serotypes. These measures will enhance the overall quality of health care and food safety, and provide better protection of the public via improved surveillance, and outbreak detection and response.

Authors' statement

All authors are members of the Canadian Public Health Laboratory Network (CPHLN) Shiga toxin-producing Escherichia coli (STEC) Working Group. This group was chaired by L Chui (Provincial Co-Chair) and S Christianson (Federal Co-Chair).

Conflict of interest

None.

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